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Matthew K. Gray

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TITLE OF THE INVENTION (250 characters max) <u>Proteotata from Plant Protoporphyrinogen Oxidase Genes</u>			

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PROMOTERS FROM PLANT PROTOPORPHYRINOGEN OXIDASE GENES

FIELD OF THE INVENTION

This invention relates to novel DNA sequences which function as promoters of transcription of associated DNA sequences in plants. More specifically, this invention relates to novel promoters which are naturally associated with plant protoporphyrinogen oxidase (protopx) coding sequences.

BACKGROUND OF THE INVENTION

I. The Protopx Enzyme and its Involvement in the Chlorophyll/Heme Biosynthetic Pathway

The biosynthetic pathways which lead to the production of chlorophyll and heme share a number of common steps. Chlorophyll is a light harvesting pigment present in all green photosynthetic organisms. Heme is a cofactor of hemoglobin, cytochromes, P450 mixed-function oxygenases, peroxidases, and catalases (see, e.g. Lehninger, *Biochemistry*, Worth Publishers, New York (1975)), and is therefore a necessary component for all aerobic organisms.

The last common step in chlorophyll and heme biosynthesis is the oxidation of protoporphyrinogen IX to protoporphyrin IX. Protoporphyrinogen oxidase (referred to herein as "protopx") is the enzyme which catalyzes this last oxidation step (Matringe *et al.*, *Biochem. J.* 260: 231 (1989)).

The protopx enzyme has been purified either partially or completely from a number of organisms including the yeast *Saccharomyces cerevisiae* (Labbe-Bois and Labbe, In *Biosynthesis of Heme and Chlorophyll*, E.H. Dailey, ed. McGraw Hill: New York, pp. 235-285 (1990)), barley etioplasts (Jacobs and Jacobs, *Biochem. J.* 244: 219 (1987)), and mouse liver (Dailey and Kart, *Biochem.* 26: 2697 (1987)). Genes encoding protopx have been isolated from two prokaryotic organisms, *Escherichia coli* (Sasarman *et al.*, *Can. J. Microbiol.* 39: 1155 (1993)) and *Bacillus subtilis* (Dailey *et al.*, *J. Biol. Chem.* 269: 813 (1994)). These genes share no sequence similarity; neither do their predicted protein products share any amino acid sequence identity. The *E. coli*

protein is approximately 21 kDa, and associates with the cell membrane. The *B. subtilis* protein is 51 kDa, and is a soluble, cytoplasmic activity.

Protox encoding cDNAs have now also been isolated from humans (see Nishimura *et al.*, *J. Biol. Chem.* 270(14): 8076-8080 (1995) and plants (International application no. PCT/IB95/00452 filed June 8, 1995, published Dec. 21, 1995 as WO 95/34659).

II. The Protox Gene as a Herbicide Target

The use of herbicides to control undesirable vegetation such as weeds or plants in crops has become almost a universal practice. The relevant market exceeds a billion dollars annually. Despite this extensive use, weed control remains a significant and costly problem for farmers.

Effective use of herbicides requires sound management. For instance, time and method of application and stage of weed plant development are critical to getting good weed control with herbicides. Since various weed species are resistant to herbicides, the production of effective herbicides becomes increasingly important.

Unfortunately, herbicides that exhibit greater potency, broader weed spectrum and more rapid degradation in soil can also have greater crop phytotoxicity. One solution applied to this problem has been to develop crops which are resistant or tolerant to herbicides. Crop hybrids or varieties resistant to the herbicides allow for the use of the herbicides without attendant risk of damage to the crop. Development of resistance can allow application of a herbicide to a crop where its use was previously precluded or limited (e.g. to pre-emergence use) due to sensitivity of the crop to the herbicide. For example, U.S. Patent No. 4,761,373 to Anderson *et al.* is directed to plants resistant to various imidazolinone or sulfonamide herbicides. The resistance is conferred by an altered acetohydroxyacid synthase (AHAS) enzyme. U.S. Patent No. 4,975,374 to Goodman *et al.* relates to plant cells and plants containing a gene encoding a mutant glutamine synthetase (GS) resistant to inhibition by herbicides that were known to inhibit GS, e.g. phosphinothricin and methionine sulfoximine. U.S. Patent No. 5,013,659 to Bedbrook *et al.* is directed to plants that express a mutant acetolactate synthase which renders the plants resistant to inhibition by sulfonylurea herbicides. U.S. Patent No. 5,162,602 to Somers *et al.* discloses plants

tolerant to inhibition by cyclohexanedione and aryloxyphenoxypropionic acid herbicides. The tolerance is conferred by an altered acetyl coenzyme A carboxylase (ACCase).

The protox enzyme serves as the target for a variety of herbicidal compounds. The herbicides that inhibit protox include many different structural classes of molecules (Duke *et al.*, *Weed Sci.* 39: 465 (1991); Nandihalli *et al.*, *Pesticide Biochem. Physiol.* 43: 193 (1992); Matringe *et al.*, *FEBS Lett.* 245: 35 (1989); Yamae and Andoh, *Pesticide Biochem. Physiol.* 35: 70 (1989)). These herbicidal compounds include the diphenylethers (e.g. acifluorfen, 5-[2-chloro-4-(trifluoromethyl)phenoxy]-2-nitrobenzoic acid; its methyl ester, or oxyfluorfen, 2-chloro-1-(3-ethoxy-4-nitrophenoxy)-4-(trifluorobenzene)), oxadiazoles (e.g. oxidiazon, 3-[2,4-dichloro-5-(1-methylethoxy)phenyl]-5-(1,1-dimethylethyl)-1,3,4-oxadiazol-2-(3H)-one), cyclic imides (e.g. S-23142, *N*-(4-chloro-2-fluoro-5-propargyloxyphenyl)-3,4,5,6-tetrahydrophthalimide; chlorophthalim, *N*-(4-chlorophenyl)-3,4,5,6-tetrahydrophthalimide), phenyl pyrazoles (e.g. TNPP-ethyl, ethyl 2-[1-(2,3,4-trichlorophenyl)-4-nitropyrzazolyl-5-oxo]propionate; M&B 39279), pyridine derivatives (e.g. LS 82-556), and phenopylate and its *O*-phenylpyrrolidino- and piperidinocarbamate analogs. Many of these compounds competitively inhibit the normal reaction catalyzed by the enzyme, apparently acting as substrate analogs.

Typically, the inhibitory effect on protox is determined by measuring fluorescence at about 622 to 635 nm, after excitation at about 395 to 410 nm (see, e.g. Jacobs and Jacobs, *Enzyme* 28: 206 (1982); Sherman *et al.*, *Plant Physiol.* 97: 280 (1991)). This assay is based on the fact that protoporphyrin IX is a fluorescent pigment, and protoporphyrinogen IX is nonfluorescent.

The predicted mode of action of protox-inhibiting herbicides involves the accumulation of protoporphyrinogen IX in the chloroplast. This accumulation is thought to lead to leakage of protoporphyrinogen IX into the cytosol where it is oxidized by a peroxidase activity to protoporphyrin IX. When exposed to light, protoporphyrin IX can cause formation of singlet oxygen in the cytosol. This singlet oxygen can in turn lead to the formation of other reactive oxygen species, which can cause lipid peroxidation and membrane disruption leading to rapid cell death (Lee *et al.*, *Plant Physiol.* 102: 881 (1993)).

Not all protox enzymes are sensitive to herbicides which inhibit plant protox enzymes. Both of the protox enzymes encoded by genes isolated from *Escherichia coli* (Sasaman *et al.*,

Can. J. Microbiol. 39: 1155 (1993)) and *Bacillus subtilis* (Dailly *et al.*, *J. Biol. Chem.* 269: 813 (1994)) are resistant to these herbicidal inhibitors. In addition, mutants of the unicellular alga *Chlamydomonas reinhardtii* resistant to the phenyltinide herbicide S-2142 have been reported (Katsuka *et al.*, *J. Pesticide Sci.* 15: 449 (1990); Shibata *et al.*, In *Research in Photosynthesis*, Vol. III, N. Murata, ed. Kluwer, Netherlands, pp. 567-570 (1992)). At least one of these mutants appears to have an altered protox activity that is resistant not only to the herbicidal inhibitor on which the mutant was selected, but also to other classes of protox inhibitors (Oshio *et al.*, *Z. Naturforsch.* 48c: 339 (1993); Sato *et al.*, In *ACS Symposium on Porphyrin Pesticides*, S. Duke, ed. ACS Press: Washington, D.C. (1994)). A mutant tobacco cell line has also been reported that is resistant to the inhibitor S-21432 (Che *et al.*, *Z. Naturforsch.* 48c: 350 (1993). In addition, modified, inhibitor-resistant forms of plant protox coding sequences have been described in international application no. PCT/IB95/00452 filed June 8, 1995, published Dec. 21, 1995 as WO 95/34659.

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III. Regulation of Protox Gene Expression

The bulk of the research related to the protox gene which has been conducted thus far has focused upon the coding sequence and modifications to this enzyme which may render it resistant to protox inhibitors. No information is available in the art with regard to the regulatory elements which control and promote the expression of protox coding sequences in plants.

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SUMMARY OF THE INVENTION

The present invention is based on the discovery that the promoter regions naturally associated with the plant protoporphyrinogen oxidase (protox) coding sequences, referred to herein generally as the "protox promoter", are useful for promoting expression of a heterologous coding sequence in a plant.

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In accordance with this discovery, the present invention provides an isolated DNA molecule comprising a plant protox promoter. The present invention further provides a chimeric

gene comprising a plant protox promoter operably linked to a heterologous coding sequence.

Plant tissue and plants containing such a chimeric gene are also provided.

In one aspect of the invention the protox promoter is used to express herbicide resistant forms of herbicide target proteins in a plant to confer tolerance to the herbicide. According to this aspect, the protox promoter may be operably linked to a coding sequence for a herbicide-resistant plant protox protein which is resistant to inhibitors of unmodified plant protox protein.

DESCRIPTION OF THE SEQUENCE LISTING

- 10 SEQ ID No. 1: DNA coding sequence for an *Arabidopsis thaliana* protox-1 protein.
- SEQ ID No. 2: *Arabidopsis thaliana* protox-1 amino acid sequence encoded by SEQ ID No. 1.
- SEQ ID No. 3: DNA coding sequence for an *Arabidopsis thaliana* protox-2 protein.
- 15 SEQ ID No. 4: *Arabidopsis thaliana* protox-2 amino acid sequence encoded by SEQ ID No. 3.
- SEQ ID No. 5: DNA coding sequence for a maize protox-1 protein.
- SEQ ID No. 6: Maize protox-1 amino acid sequence encoded by SEQ ID No. 5.
- 20 SEQ ID No. 7: DNA coding sequence for a maize protox-2 protein.
- SEQ ID No. 8: Maize protox-2 amino acid sequence encoded by SEQ ID No. 7.
- SEQ ID No. 9: DNA coding sequence for a wheat protox-1 protein.
- SEQ ID No. 10: Wheat protox-1 amino acid sequence encoded by SEQ ID No. 9.
- SEQ ID No. 11: DNA coding sequence for a soybean protox-1 protein.
- 25 SEQ ID No. 12: Soybean protox-1 protein encoded by SEQ ID No. 11.
- SEQ ID NO. 13: Promoter sequence from *Arabidopsis thaliana* protox-1 gene.

DEFINITIONS

As used herein a "plant protox promoter" is used to refer to the regulatory region which naturally occurs immediately upstream of a protoporphyrinogen oxidase (protox) coding sequence in a plant and is responsible, in its naturally occurring state, for regulating the transcription of the associated protox coding sequence. The plant protox promoter includes the DNA region directly involved in binding of RNA polymerase to initiate transcription and additional upstream regulatory cis-elements which influence the transcription of an operably linked coding sequence.

As used herein a "gene" is used to refer to a DNA molecule which includes (1) a coding sequence and (2) associated regulatory regions which promote and regulate the transcription of the coding sequence in a suitable host cell. The coding sequence may encode a useful transcript (e.g. antisense RNA) or polypeptide produced by translation of the encoded transcript. A gene includes at a minimum, in 5'-3' orientation, a promoter region, a coding sequence and a transcription terminator. A gene may also include additional regulatory regions which can occur as part of the minimal elements (e.g. leaders or signal peptides within the coding sequence) or as discrete elements (e.g. introns).

As used herein a "chimeric gene" refers to a gene which does not naturally occur wherein at least one component part is heterologous with respect to another component part. As used herein to describe the present invention a "chimeric gene" refers to a gene which includes the promoter of the invention operably linked to a heterologous coding sequence.

As used herein with reference to the relationship between a promoter and a coding sequence, the term "heterologous" is used to refer to a relationship which does not naturally occur. For instance, a coding sequence is considered heterologous with respect to a promoter sequence if it is different from the coding sequence that naturally occurs in association with the promoter sequence. This includes modified forms of coding sequences which are naturally associated with a subject promoter. Accordingly, a modified, inhibitor-resistant protox coding sequence is considered to be heterologous with respect to the promoter that is naturally associated with the unmodified, inhibitor-sensitive form of this coding sequence.

As used herein, the term "substantial sequence homology" is used to indicate that a nucleotide sequence (in the case of DNA or RNA) or an amino acid sequence (in the case of a protein or polypeptide) exhibits substantial structural and functional equivalence with another nucleotide or amino acid sequence. Any functional or structural differences between sequences having substantial sequence homology will be de minimis; that is they will not affect the ability of the sequence to function as indicated in the present application. For example, a sequence which has substantial sequence homology with a DNA sequence disclosed to be a plant protox promoter will be able to direct the same level and pattern of expression of an associated DNA sequence as the plant protox promoter. Sequences that have substantial sequence homology with the sequences disclosed herein are usually variants of the disclosed sequence, such as mutations, but may also be synthetic sequences. Structural differences are considered de minimis if there is a significant amount of sequence overlap or similarity between two or more different sequences or if the different sequences exhibit similar physical characteristics. Such characteristics can include, for example, immunological reactivity, enzyme activity, structural protein integrity, etc.

Two nucleotide sequences may have substantial sequence homology if the sequences have at least 70 percent, more preferably 80 percent and most preferably 90 percent sequence similarity between them. Two amino acid sequences have substantial sequence homology if they have at least 50 percent, preferably 70 percent, and most preferably 90 percent similarity between the active portions of the polypeptides. In the case of promoter DNA sequences, "substantial sequence homology" also refers to those fragments of a promoter DNA sequence that are able to operate to promote the expression of associated DNA sequences. Such operable fragments of a promoter DNA sequence may be derived from the promoter DNA sequence, for example, by cleaving the promoter DNA sequence using restriction enzymes, synthesizing in accordance with the sequence of the promoter DNA sequence, or may be obtained through the use of PCR technology. Mullis et al., *Meth. Enzymol.*, 155:335-350 (1987); Erlich (ed.), *PCR Technology*. Stockton Press (New York 1989).

A promoter DNA sequence is said to be "operably linked" to a second DNA sequence if the two are situated such that the promoter DNA sequence influences the transcription or translation of the second DNA sequence. For example, if the second DNA sequence codes for

the production of a protein, the promoter DNA sequence would be operably linked to the second DNA sequence if the promoter DNA sequence affects the expression of the protein product from the second DNA sequence. For example, in a DNA sequence comprising a promoter DNA sequence physically attached to a coding DNA sequence in the same chimeric construct, the two sequences are likely to be operably linked.

DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to promoter DNA sequences which are naturally associated with coding sequences for plant protoporphyrinogen oxidase (referred to herein as "protox"; see international application no. PCT/IB95/00452 filed June 8, 1995, published Dec. 21, 1995 as WO 95/34659 and co-pending provisional application entitled "DNA Molecules Encoding Plant Protoporphyrinogen Oxidase and Inhibitor Resistant Mutants Thereof" filed on the same day as the instant application). These protox promoter sequences have been found to be useful for the expression of a heterologous coding sequence in a plant.

The promoter sequence for the *Arabidopsis thaliana* protox-1 coding sequence (SEQ ID No. 1) is provided as SEQ ID No. 13. Isolation of this promoter from a genomic library using the associated coding sequence as a probe is described in Example 1. This same approach can be used to isolate the promoter sequence from any plant protox gene. Any protox coding sequence which shares sufficient homology to hybridize to the protox coding sequence associated with the promoter of interest may be used as a probe in this approach. Since the respective protox-1 and protox-2 coding sequences from all plants are contemplated to share this requisite degree of homology, the choice of which protox coding sequence is used as a probe is not considered critical. However, for optimal hybridization results it is preferable to use the most closely related protox coding sequence. Most preferably, the coding sequence used as a probe is from the same plant species as the protox promoter of interest and is the coding sequence naturally associated with the promoter.

The plant protox promoter of the present invention includes the *Arabidopsis* protox-1 promoter sequence set forth in SEQ ID No. 13 as well as corresponding protox-1 promoter sequences available from other plant species as indicated above. The present invention also

the production of a protein, the promoter DNA sequence would be operably linked to the second DNA sequence if the promoter DNA sequence affects the expression of the protein product from the second DNA sequence. For example, in a DNA sequence comprising a promoter DNA sequence physically attached to a coding DNA sequence in the same chimeric construct, the two sequences are likely to be operably linked.

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The plant protox promoter of the present invention includes the *Arabidopsis* protox-1 promoter sequence set forth in SEQ ID No. 13 as well as corresponding protox-1 promoter sequences available from other plant species as indicated above. The present invention also

includes functional fragments of these DNA sequences which retain the ability to regulate expression of an operably linked coding sequence in the same manner as the exemplified protox promoter sequence. Such functional fragments may be identified through deletion analyses or other standard techniques used in the art to identify protox promoter activity (see, e.g. pages 546-549 of "Genes IV", ed. by Lewin, Oxford Univ. Press (1990)). The present invention also includes DNA sequences having substantial sequence homology with the protox promoters available from plant genes which confer an equivalent level and pattern of expression upon an operably linked sequence. Such promoter sequences may be obtained through modification of the protox promoters isolated from plant genes and are considered functionally equivalent derivatives of the plant protox promoters.

As illustrated in the examples below, the DNA sequences, vectors and transgenic plants of the present invention comprise a promoter sequence derived from a plant protox gene. The protox promoter DNA sequences are preferably linked operably to a coding DNA sequence, for example a DNA sequence which is transcribed into a useful RNA transcript such as an antisense transcript, or a coding sequence which is ultimately expressed in the production of a useful protein product.

In a preferred embodiment, the protox promoter is used to direct the expression of a modified herbicide target enzyme which is resistant to herbicides at levels that inhibit the corresponding unmodified version of the enzyme. Such modified herbicide-resistant enzymes include herbicide-resistant forms of imidazoglycerol phosphate dehydratase (IGPD; see WO 9426909 published Nov. 24, 1994), EPSP synthase (see U.S. Pat. Nos. 4,535,060; 4,769,061; 4,940,835 and EP 550,633), glutamine synthetase (GS; see U.S. Patent No. 4,975,374), acetyl coenzyme A carboxylase (ACCase; see U.S. Patent No. 5,162,602), and acetolactate synthase (see U.S. Patent Nos. 4,761,373; 5,304,732; 5,331,107; 5,013,659; 5,141,870; and 5,378,824).

In a most preferred embodiment, the protox promoter is used to direct the expression of a modified protox enzyme which is resistant to protox inhibitors as illustrated in Examples 2-3 (see also International application no. PCT/IB95/00452 filed June 8, 1995, published Dec. 21, 1995 as WO 95/34659 whose relevant parts are herein incorporated by reference; see also co-pending

application entitled "DNA Molecules Encoding Plant Protoporphyrinogen Oxidase and Inhibitor-Resistant Mutants Thereof" filed on the same day as the instant application).

The transgenic plants of the present invention may be transformed by any method of transformation known in the art. These methods include, for instance, transformation by direct infection or co-cultivation of plants, plant tissue or cells with *Agrobacterium tumefaciens*; Horsc
 5 *et al.*, *Science*, 225: 1229 (1985); Marton, "Cell Culture and Somatic Cell Genetic of Plants", vol. 1, pp 514-521 (1984); direct gene transfer into protoplasts; Paszkowski *et al.*, *EMBO J.* 12: 2717 (1984); Loez *et al.*, *Mol. Gen. & Genet.* 1199:178 (1985); Fromm *et al.*, *Nature* 319:719 (1986); microprojectile bombardment, Klein *et al.*, *BioTechnology*, 6:559-563 (1988); injection into
 10 protoplasts cultured cells and tissues, Reich *et al.*, *BioTechnology*, 4:1001-1004 (1986); or injection into meristematic tissues of seedlings and plants as described by De La Pena *et al.*, *Nature*, 325:274-276 (1987); Hooykaas-Van Slooten *et al.*, *Nature*, 311:763-764 (1984); Grimalley *et al.*, *BioTechnology*, 6:185 (1988); and Grimalley *et al.*, *Nature*, 325:177 (1988).

The invention is illustrated in more detail by the following examples, without implying
 15 any restriction to what is described therein.

EXAMPLES

EXAMPLE 1: Isolation of the *Arabidopsis thaliana* Proton-1 promoter sequence

20 A Lambda Zap II genomic DNA library prepared from *Arabidopsis thaliana* (Columbia, whole plant) was purchased from Stratagene. Approximately 125,000 phage were plated at a density of 25,000 pfu (plaque forming units) per 15 cm Petri dish and duplicate lifts were made onto Colony/Plaque Screen membranes (NEN Dupont). The plaque lifts were probed with the *Arabidopsis* Proton-1 cDNA (SEQ ID No. 1) labeled with 32P-dCTP by the random priming
 25 method (Life Technologies). Hybridization and wash conditions were at 65° C as described in Church and Gilbert, *Proc. Natl. Acad. Sci. USA* 81: 1991-1995 (1984). Positively hybridizing plaques were purified and *in vivo* excised into pBluescript plasmids. Sequence from the genomic DNA inserts was determined by the chain termination method using dideoxy terminators labeled with fluorescent dyes (Applied Biosystems, Inc.). One clone, AraPT1Pro, was determined to
 30 contain 580 bp of *Arabidopsis* sequence upstream from the initiating methionine (ATG) of the

Protox-1 protein coding sequence. This clone also contains coding sequence and introns that extend to bp 1241 of the Protox-1 cDNA sequence. The 580 bp 5' noncoding fragment is the putative Arabidopsis Protox-1 promoter, and the sequence is set forth in SEQ ID No. 13.

AraPT1Pro was deposited December 14, 1995, as pWDC-11 (NRRL #B-21515)

EXAMPLE 2: Construction of plant transformation vectors expressing altered Protox-1 genes behind the native Arabidopsis Protox-1 promoter

A full-length cDNA of the appropriate altered Arabidopsis Protox-1 cDNA is isolated as an EcoRI-XhoI partial digest fragment and cloned into the plant expression vector pCGN1761ENX (see Example 9 of International application no. PCT/IB95/00452 filed June 8, 1995, published Dec. 21, 1995 as WO 95/34659). This plasmid is digested with NcoI and BamHI to produce a fragment comprised of the complete Protox-1 cDNA plus a transcription terminator from the 3' untranslated sequence of the trnI gene of *Agrobacterium tumefaciens*. The AraPT1Pro plasmid described above is digested with NcoI and BamHI to produce a fragment comprised of pBluescript and the 580 bp putative Arabidopsis Protox-1 promoter. Ligation of these two fragments produces a fusion of the altered protox cDNA to the native protox promoter. The expression cassette containing the Protox-1 promoter/Protox-1 cDNA/trnI terminator fusion is excised by digestion with KpnI and cloned into the binary vector pCIB200. The binary plasmid is transformed by electroporation into *Agrobacterium* and then into *Arabidopsis* using the vacuum infiltration method (Bechtold *et al. C.R. Acad. Sci. Paris* 316: 1194-1199 (1993)). Transformants expressing altered protox genes are selected on kanamycin or on various concentrations of protox inhibiting herbicide.

EXAMPLE 3: Production of herbicide tolerant plants by expression of a native Protox-1 promoter/alternated Protox-1 fusion

Using the procedure described above, an Arabidopsis Protox-1 cDNA containing a TAC to ATG (Tyrosine to Methionine) change at nucleotides 1306-1308 in the Protox-1 sequence (SEQ ID No.1) was fused to the native Protox-1 promoter fragment and transformed into *Arabidopsis thaliana*. This altered Protox-1 enzyme (AraC-2Met) has been shown to be >10fold

more tolerant to various protox-inhibiting herbicides than the naturally occurring enzyme when tested in a bacterial expression system (see Example 5 of copending U.S. application entitled "DNA Molecules Encoding Plant Protoporphyrinogen Oxidase and Inhibitor Resistant Mutants Thereof" filed on the same day as the instant application). Seed from the vacuum infiltrated plants was collected and plated on a range (10.0nM-1.0uM) of a protox inhibitory aryloxyacetyl herbicide of formula XVII. Multiple experiments with wild type *Arabidopsis* have shown that a 10.0nM concentration of this compound is sufficient to prevent normal seedling germination. Transgenic seeds expressing the AraC-2Met altered enzyme fused to the native Protop-1 promoter produced normal *Arabidopsis* seedlings at herbicide concentrations up to 500nM, indicating at least 50-fold higher herbicide tolerance when compared to wild-type *Arabidopsis*. This promoter/alterd protox enzyme fusion therefore functions as an effective selectable marker for plant transformation. Several of the plants that germinated on 100.0nM of protox-inhibiting herbicide were transplanted to soil, grown 2-3 weeks, and tested in a spray assay with various concentrations of the protox-inhibiting herbicide. When compared to empty vector control transformants, the AraPT1Pro/AraC-2Met transgenics were >10fold more tolerant to the herbicide spray.

EXAMPLE 4: Isolation of Maize Protop-1 promoter sequences

A *Zea mays* (Missouri 17 inbred, etiolated seedlings) genomic DNA library in the Lambda FIX II vector was purchased from Stratagene. Approximately 250,000 pfu of the library was plated at a density of 50,000 phage per 15 cm plate and duplicate lifts were made onto Colony/Plaque screen membranes (NEN Dupont). The plaque lifts were probed with the maize Protop-1 cDNA (SEQ ID No. 5) labeled with 32P-dCTP by the random priming method (Life Technologies). Hybridization and wash conditions were at 65° C as described in Church and Gilbert, *Proc. Natl. Acad. Sci. USA* 81: 1991-1995 (1984). Positively hybridizing plaques were purified and rescreened with a 210 bp EcoRI-NcoI fragment from the 5' end of the maize Protop-1 cDNA. Lambda phage DNA was isolated from three phage that hybridized to the 5' fragment using the Wizard Lambda Preps DNA Purification System (Promega). Restriction analysis and hybridization to the 5' maize fragment indicated that two of the phage clones are derived from the

same gene, while a third may represent a second unique *Protoc-1* gene. Hybridizing fragments from both types of phage are subcloned into a pBluecript vector for sequence analysis.

EXAMPLE 5: Construction of Plant Transformation Vectors

5 Numerous transformation vectors are available for plant transformation, and the promoters and chimeric genes of this invention can be used in conjunction with any such vectors. The selection of vector for use will depend upon the preferred transformation techniques and the target species for transformation. For certain target species, different antibiotic or herbicide selection markers may be preferred. Selection markers used routinely in transformation include
10 the *aptII* gene which confers resistance to kanamycin and related antibiotics (Messing & Vieira, *Gene* 19: 259-268 (1982); Bevan *et al.*, *Nature* 304: 184-187 (1983)), the *bar* gene which confers resistance to the herbicide phosphinothricin (White *et al.*, *Nucl Acids Res* 18: 1062 (1990), Spencer *et al.* *Theor Appl Genet* 79: 625-631 (1990)), the *hph* gene which confers resistance to the antibiotic hygromycin (Blochinger & Diggelmann, *Mol Cell Biol* 4: 2929-2931), and the *dhfr*
15 gene, which confers resistance to methotrexate (Bourouis *et al.*, *EMBO J.* 2(7): 1099-1104 (1983)).

(1) Construction of Vectors Suitable for *Agrobacterium* Transformation

Many vectors are available for transformation using *Agrobacterium tumefaciens*. These typically
20 carry at least one T-DNA border sequence and include vectors such as pBIN19 (Bevan, *Nucl Acids Res.* (1984)) and pXYZ. Below the construction of two typical vectors is described.

Construction of pCIB200 and pCIB2001

The binary vectors pCIB200 and pCIB2001 are used for the construction of recombinant
25 vectors for use with *Agrobacterium* and was constructed in the following manner. pTJS75kan was created by *NarI* digestion of pTJS75 (Schmidhauser & Helinski, *J Bacteriol.* 164: 446-455

(1985)) allowing excision of the tetracycline-resistance gene, followed by insertion of an *AccI* fragment from pUC4K carrying an NPTII (Messing & Vieira, *Gene* 19: 259-268 (1982); Bevan *et al.*, *Nature* 304: 184-187 (1983); McBride *et al.*, *Plant Molecular Biology* 14: 266-276 (1990)). *XhoI* linkers were ligated to the *EcoRV* fragment of pCIB7 which contains the left and right T-DNA borders, a plant selectable *nos/nptII* chimeric gene and the pUC polylinker (Rothstein *et al.*, *Gene* 53: 153-161 (1987)), and the *XhoI*-digested fragment was cloned into *SacI*-digested pTIS75kan to create pCIB200 (see also EP 0 332 104, example 19 [1338]). pCIB200 contains the following unique polylinker restriction sites: *EcoRI*, *SacI*, *KpnI*, *BglII*, *XbaI*, and *Sall*. pCIB2001 is a derivative of pCIB200 which created by the insertion into the polylinker of additional restriction sites. Unique restriction sites in the polylinker of pCIB2001 are *EcoRI*, *SacI*, *KpnI*, *BglII*, *XbaI*, *Sall*, *MluI*, *BclI*, *AvrII*, *ApaI*, *HpaI*, and *SnaI*. pCIB2001, in addition to containing these unique restriction sites also has plant and bacterial kanamycin selection, left and right T-DNA borders for *Agrobacterium*-mediated transformation, the RK2-derived *trfA* function for mobilization between *E. coli* and other hosts, and the *oriT* and *oriV* functions also from RK2. The pCIB2001 polylinker is suitable for the cloning of plant expression cassettes containing their own regulatory signals.

Construction of pCIB10 and Hygromycin Selection Derivatives thereof

The binary vector pCIB10 contains a gene encoding kanamycin resistance for selection in plants, T-DNA right and left border sequences and incorporates sequences from the wide host-range plasmid pRK252 allowing it to replicate in both *E. coli* and *Agrobacterium*. Its construction is described by Rothstein *et al.*, *Gene* 53: 153-161 (1987). Various derivatives of pCIB10 have been constructed which incorporate the gene for hygromycin B phosphotransferase described by Gritz *et al.*, *Gene* 25: 179-188 (1983)). These derivatives enable selection of transgenic plant cells on hygromycin only (pCIB743), or hygromycin and kanamycin (pCIB715, pCIB717).

(2) Construction of Vectors Suitable for non-*Agrobacterium* Transformation.

Transformation without the use of *Agrobacterium tumefaciens* circumvents the requirement for T-DNA sequences in the chosen transformation vector and consequently vectors lacking these sequences can be utilized in addition to vectors such as the ones described above which contain T-DNA sequences. Transformation techniques which do not rely on *Agrobacterium* include transformation via particle bombardment, protoplast uptake (e.g. PEG and electroporation) and microinjection. The choice of vector depends largely on the preferred selection for the species being transformed. Below, the construction of some typical vectors is described.

Construction of pCIB3064

pCIB3064 is a pUC-derived vector suitable for direct gene transfer techniques in combination with selection by the herbicide basta (or phosphinothricin). The plasmid pCIB246 comprises the CaMV 35S promoter in operational fusion to the *E. coli* GUS gene and the CaMV 35S transcriptional terminator and is described in the PCT published application WO 93/07278. The 35S promoter of this vector contains two ATG sequences 5' of the start site. These sites were mutated using standard PCR techniques in such a way as to remove the ATGs and generate the restriction sites *SapI* and *PvuII*. The new restriction sites were 96 and 37 bp away from the unique *SalI* site and 101 and 42 bp away from the actual start site. The resultant derivative of pCIB246 was designated pCIB3025. The GUS gene was then excised from pCIB3025 by digestion with *SalI* and *SacI*, the termini rendered blunt and religated to generate plasmid pCIB3060. The plasmid pJIT82 was obtained from the John Innes Centre, Norwich and the 400 bp *SmaI* fragment containing the *bar* gene from *Streptomyces viridochromogenes* was excised and inserted into the *HpaI* site of pCIB3060 (Thompson *et al.* EMBO J 6: 2519-2523 (1987)). This generated pCIB3064 which comprises the *bar* gene under the control of the CaMV 35S promoter and terminator for herbicide selection, a gene for ampicillin resistance (for selection in

E. coli) and a polylinker with the unique sites *SphI*, *PstI*, *HindIII*, and *BamHI*. This vector is suitable for the cloning of plant expression cassettes containing their own regulatory signals.

Construction of pSOG19 and pSOG35

5 pSOG35 is a transformation vector which utilizes the *E. coli* gene dihydrofolate reductase (DHFR) as a selectable marker conferring resistance to methotrexate. PCR was used to amplify the 35S promoter (~800 bp), intron 6 from the maize *Adh1* gene (~550 bp) and 18 bp of the GUS untranslated leader sequence from pSOG10. A 250 bp fragment encoding the *E. coli* dihydrofolate reductase type II gene was also amplified by PCR and these two PCR fragments
10 were assembled with a *SacI*-*PstI* fragment from pBI221 (Clontech) which comprised the pUC19 vector backbone and the nopaline synthase terminator. Assembly of these fragments generated pSOG19 which contains the 35S promoter in fusion with the intron 6 sequence, the GUS leader, the DHFR gene and the nopaline synthase terminator. Replacement of the GUS leader in
15 pSOG19 with the leader sequence from Maize Chlorotic Mottle Virus (MCMV) generated the vector pSOG35. pSOG19 and pSOG35 carry the pUC gene for ampicillin resistance and have *HindIII*, *SphI*, *PstI* and *EcoRI* sites available for the cloning of foreign sequences such as chimeric gene sequences containing a plant protox promoter.

EXAMPLE 12: Construction of Chimeric Genes/Plant Expression Cassettes

20 Coding sequences intended for expression in transgenic plants under the control of a plant protox promoter may be assembled in expression cassettes behind a suitable protox promoter and upstream of a suitable transcription terminator. The resulting chimeric genes can then be easily transferred to the plant transformation vectors described above in Example 19.

25 Protox Promoter Selection

In accordance with the present invention, the chimeric gene will contain a plant protox promoter. The selection of the specific protox promoter used in the chimeric gene is primarily up

to the individual researcher, although generally it will be preferable to use a protox promoter from a plant species closely related to, or most preferably identical, to the species intended to contain the resulting chimeric gene. For example, if the chimeric gene is intended to be contained in a maize plant it would be preferable to use a protox promoter from a monocotyledonous plant and most preferable to use a maize protox promoter.

Transcriptional Terminators

A variety of transcriptional terminators are available for use in expression cassettes. These are responsible for the termination of transcription beyond the transgene and its correct polyadenylation. Appropriate transcriptional terminators are those which are known to function in plants and include the CaMV 35S terminator, the *sv4* terminator, the sopaline synthase terminator, the pea *rbcS* E9 terminator, as well as terminators naturally associated with the plant protox gene (i.e. "prototox terminators"). These can be used in both monocotyledons and dicotyledons.

Sequences for the Enhancement or Regulation of Expression

Numerous sequences have been found to enhance gene expression from within the transcriptional unit and these sequences can be used in conjunction with the genes of this invention to increase their expression in transgenic plants.

Various intron sequences have been shown to enhance expression, particularly in monocotyledonous cells. For example, the introns of the maize *Adhl* gene have been found to significantly enhance the expression of the wild-type gene under its cognate promoter when introduced into maize cells. Intron 1 was found to be particularly effective and enhanced expression in fusion constructs with the chloramphenicol acetyltransferase gene (Callis *et al.*, Genes Develop. 1: 1183-1200 (1987)). In the same experimental system, the intron from the maize *bronze1* gene had a similar effect in enhancing expression (Callis *et al.*, *agru*). Intron sequences have been routinely incorporated into plant transformation vectors, typically within the non-translated leader.

A number of non-translated leader sequences derived from viruses are also known to enhance expression, and these are particularly effective in dicotyledonous cells. Specifically, leader sequences from Tobacco Mosaic Virus (TMV, the "W-sequence"), Maize Chlorotic Mottle Virus (MCMV), and Alfalfa Mosaic Virus (AMV) have been shown to be effective in enhancing
5 expression (e.g. Gallie *et al. Nucl. Acids Res.* 15: 8693-8711 (1987); Skuzeski *et al. Plant Molec. Biol.* 15: 65-79 (1990))

Targeting of the Gene Product Within the Cell

Various mechanisms for targeting gene products are known to exist in plants and the
10 sequences controlling the functioning of these mechanisms have been characterized in some detail. For example, the targeting of gene products to the chloroplast is controlled by a signal sequence found at the amino terminal end of various proteins and which is cleaved during chloroplast import yielding the mature protein (e.g. Comai *et al. J. Biol. Chem.* 263: 15104-15109 (1988)).
These signal sequences can be fused to heterologous gene products to effect the import of
15 heterologous products into the chloroplast (van den Broeck *et al. Nature* 313: 358-363 (1985)). DNA encoding for appropriate signal sequences can be isolated from the 5' end of the cDNAs encoding the RUBISCO protein, the CAB protein, the EPSP synthase enzyme, the GS2 protein and many other proteins which are known to be chloroplast localized.

Other gene products are localized to other organelles such as the mitochondrion and the
20 peroxisome (e.g. Unger *et al. Plant Molec. Biol.* 13: 411-418 (1989)). The cDNAs encoding these products can also be manipulated to effect the targeting of heterologous gene products to these organelles. Examples of such sequences are the nuclear-encoded ATPases and specific aspartate amino transferase isoforms for mitochondria. Targeting to cellular protein bodies has been described by Rogers *et al., Proc. Natl. Acad. Sci. USA* 82: 6512-6516 (1985)).

25 In addition sequences have been characterized which cause the targeting of gene products to other cell compartments. Amino terminal sequences are responsible for targeting to the ER, the apoplast, and extracellular secretion from aleurone cells (Koehler & Ho, *Plant Cell* 2: 769-783 (1990)). Additionally, amino terminal sequences in conjunction with carboxy terminal

sequences are responsible for vacuolar targeting of gene products (Shinshi *et al.*, *Plant Molec. Biol.* 14: 357-368 (1990)).

By the fusion of the appropriate targeting sequences described above to transgene sequences of interest it is possible to direct the transgene product to any organelle or cell compartment. For chloroplast targeting, for example, the chloroplast signal sequence from the RUBISCO gene, the CAB gene, the EPSP synthase gene, or the GS2 gene is fused in frame to the amino terminal ATG of the transgene. The signal sequence selected should include the known cleavage site and the fusion constructed should take into account any amino acids after the cleavage site which are required for cleavage. In some cases this requirement may be fulfilled by the addition of a small number of amino acids between the cleavage site and the transgene ATG or alternatively replacement of some amino acids within the transgene sequence. Posions constructed for chloroplast import can be tested for efficacy of chloroplast uptake by *in vitro* translation of *in vitro* transcribed constructions followed by *in vitro* chloroplast uptake using techniques described by (Bartlett *et al.* In: Edelman *et al.* (Eds.) *Methods in Chloroplast Molecular Biology*, Elsevier, pp 1081-1091 (1982); Wassmann *et al.* *Mol. Gen. Genet.* 205: 446-453 (1986)). These construction techniques are well known in the art and are equally applicable to mitochondria and peroxisomes. The choice of targeting which may be required for expression of the transgenes will depend on the cellular localization of the precursor required as the starting point for a given pathway. This will usually be cytosolic or chloroplastic, although it may in some cases be mitochondrial or peroxisomal. The products of transgene expression will not normally require targeting to the ER, the apoplast or the vacuole.

The above described mechanisms for cellular targeting can be utilized in conjunction with plant protox promoters so as to effect a specific cell targeting goal under the transcriptional regulation of a promoter which has an expression pattern different to that of the promoter from which the targeting signal derives.

EXAMPLE 13: Transformation of Dicotyledons

Transformation techniques for dicotyledons are well known in the art and include *Agrobacterium*-based techniques and techniques which do not require *Agrobacterium*. Non-

Agrobacterium techniques involve the uptake of exogenous genetic material directly by protoplasts or cells. This can be accomplished by PEG or electroporation mediated uptake, particle bombardment-mediated delivery, or microinjection. Examples of these techniques are described by Paszkowski *et al.*, *EMBO J* 3: 2717-2722 (1984), Potrykus *et al.*, *Mol. Gen. Genet.* 199: 169-177 (1985), Reich *et al.*, *Biotechnology* 4: 1001-1004 (1986), and Klein *et al.*, *Nature* 327: 70-73 (1987). In each case the transformed cells are regenerated to whole plants using standard techniques known in the art.

Agrobacterium-mediated transformation is a preferred technique for transformation of dicotyledons because of its high efficiency of transformation and its broad utility with many different species. The many crop species which are routinely transformable by *Agrobacterium* include tobacco, tomato, sunflower, cotton, oilseed rape, potato, soybean, alfalfa and poplar (EP 0 317 511 (cotton), EP 0 249 432 (tomato, to Calgene), WO 87/07299 (*Brassica*, to Calgene), US 4,795,855 (poplar)).

Transformation of the target plant species by recombinant *Agrobacterium* usually involves co-cultivation of the *Agrobacterium* with explants from the plant and follows protocols well known in the art. Transformed tissue is regenerated on selectable medium carrying the antibiotic or herbicide resistance marker present between the binary plasmid T-DNA borders.

EXAMPLE 14: Transformation of Monocotyledons

Transformation of most monocotyledon species has now also become routine. Preferred techniques include direct gene transfer into protoplasts using PEG or electroporation techniques, and particle bombardment into callus tissue. Transformations can be undertaken with a single DNA species or multiple DNA species (*i.e.* co-transformation) and both these techniques are suitable for use with this invention. Co-transformation may have the advantage of avoiding complex vector construction and of generating transgenic plants with unlinked loci for the gene of interest and the selectable marker, enabling the removal of the selectable marker in subsequent generations, should this be regarded desirable. However, a disadvantage of the use of co-transformation is the less than 100% frequency with which separate DNA species are integrated into the genome (Schocher *et al.* *Biotechnology* 4: 1093-1096 (1986)).

Patent Applications EP 0 292 435 (to Ciba-Geigy), EP 0 392 225 (to Ciba-Geigy), WO 93/07278 (to Ciba-Geigy) and U.S. Patent No. 5,350,689 (to Ciba-Geigy) describe techniques for the preparation of callus and protoplasts from an elite inbred line of maize, transformation of protoplasts using PEG or electroporation, and the regeneration of maize plants from transformed
5 protoplasts. Gordon-Kamm *et al.*, *Plant Cell* 2: 603-618 (1990)) and Fromm *et al.*, *Biotechnology* 8: 833-839 (1990)) have published techniques for transformation of A188-derived maize line using particle bombardment. Furthermore, application WO 93/07278 (to Ciba-Geigy) and Kotiel *et al.*, *Biotechnology* 11: 194-200 (1993)) describe techniques for the transformation of elite inbred lines of maize by particle bombardment. This technique utilizes immature maize
10 embryos of 1.5-2.5 mm length excised from a maize ear 14-15 days after pollination and a PDS-1000E Biolistica device for bombardment.

Transformation of rice can also be undertaken by direct gene transfer techniques utilizing protoplasts or particle bombardment. Protoplast-mediated transformation has been described for Japonica-types and Indica-types (Zhang *et al.*, *Plant Cell Rep* 7: 379-384 (1988); Shimamoto *et al.* *Nature* 338: 274-277 (1989); Datta *et al.* *Biotechnology* 8: 736-740 (1990)). Both types are
15 also routinely transformable using particle bombardment (Christou *et al.* *Biotechnology* 9: 957-962 (1991)).

Patent Application EP 0 332 581 (to Ciba-Geigy) describes techniques for the generation, transformation and regeneration of Poaceae protoplasts. These techniques allow the
20 transformation of *Dactylis* and wheat. Furthermore, wheat transformation was been described by Vasil *et al.*, *Biotechnology* 10: 667-674 (1992)) using particle bombardment into cells of type C long-term regenerable callus, and also by Vasil *et al.*, *Biotechnology* 11: 1553-1558 (1993)) and Weeks *et al.*, *Plant Physiol.* 102: 1077-1084 (1993) using particle bombardment of immature embryos and immature embryo-derived callus. A preferred technique for wheat transformation,
25 however, involves the transformation of wheat by particle bombardment of immature embryos and includes either a high sucrose or a high maltose step prior to gene delivery. Prior to bombardment, any number of embryos (0.75-1 mm in length) are plated onto MS medium with 3% sucrose (Murashige & Skoog, *Physiologia Plantarum* 15: 473-497 (1962)) and 3 mg/l 2,4-D for induction of somatic embryos which is allowed to proceed in the dark. On the chosen day of

5 bombardment, embryos are removed from the induction medium and placed onto the osmoticum
(i.e. induction medium with sucrose or maltose added at the desired concentration, typically
15%). The embryos are allowed to plasmolyze for 2-3 h and are then bombarded. Twenty
embryos per target plate is typical, although not critical. An appropriate gene-carrying plasmid
5 (such as pCIB3064 or pSG35) is precipitated onto micrometer size gold particles using standard
procedures. Each plate of embryos is shot with the DuPont Biotistics® helium device using a burst
pressure of ~1000 psi using a standard 80 mesh screen. After bombardment, the embryos are
placed back into the dark to recover for about 24 h (still on osmoticum). After 24 hrs, the
embryos are removed from the osmoticum and placed back onto induction medium where they
10 stay for about a month before regeneration. Approximately one month later the embryo explants
with developing embryogenic callus are transferred to regeneration medium (MS + 1 mg/liter
NAA, 5 mg/liter GA), further containing the appropriate selection agent (10 mg/l basta in the case
of pCIB3064 and 2 mg/l methotrexate in the case of pSG35). After approximately one month,
developed shoots are transferred to larger sterile containers known as "GA7s" which contained
15 half-strength MS, 2% sucrose, and the same concentration of selection agent. Patent application
08/147,161 describes methods for wheat transformation and is hereby incorporated by reference.

20

While the present invention has been described with reference to specific embodiments
thereof, it will be appreciated that numerous variations, modifications, and embodiments are
possible, and accordingly, all such variations, modifications and embodiments are to be regarded
as being within the spirit and scope of the present invention.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

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(ii) TITLE OF INVENTION: PROMOTERS FROM PLANT PROTOPORPHYRINOGEN
OXIDASE GENES

(iii) NUMBER OF SEQUENCES: 13

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(v) COMPUTER READABLE FORM:

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(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

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(A) APPLICATION NUMBER: US TBA
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(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1719 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 31..1644

(D) OTHER INFORMATION: /note= "Arabidopsis protein-1 CDS;
sequence from pMDC-2"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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ACG ACT CAA TCG CTT CTT CCG TCG TTT TCG AAG CCG AAT CTC CGA TTA      102
Thr Thr Gln Ser Leu Leu Pro Ser Phe Ser Lys Pro Asn Leu Arg Leu
10 15 20

AAT GTT TAT AAG CCT CTT AGA CTC CDT TGT TCA GTG CCG GGT GGA CCA      150
Asn Val Tyr Lys Pro Leu Arg Leu Arg Cys Ser Val Ala Gly Gly Pro
25 30 35 40

ACC GTC CGA TCT TCA AAA ATC GAA GCG GGA CGA GCG ACC ACC ATC ACC      198
Thr Val Gly Ser Ser Lys Ile Glu Gly Gly Gly Gly Thr Thr Ile Thr
45 50 55

ACG GAT TGT GTG ATT GTC GCG GGA GGT AAT AAT GGT CTT TGC ATC GCT      246
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75 80 85

ACC GAG GCT AAG GAT CGT GTT GGA GCG AAC ATT ATC ACT CDT GAA GAG      342
Thr Glu Ala Lys Asp Arg Val Gly Gly Asn Ile Ile Thr Arg Glu Glu
90 95 100

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105 110 115 120

CCT ATG CTC ACT ATG GTG GTA GAT AAT GGT TTC AAG GAT GAT TTG GTG      438
Pro Met Leu Thr Met Val Val Asp Ser Gly Leu Lys Asp Asp Leu Val
125 130 135

TTG GGA GAT CTT ACT CCG CCA AGG TTT GTG TTG TGG AAT GCG AAA TTG      486
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140 145 150

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24

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Lys	Gly	Leu	Arg	Met	Leu	Pro	Glu	Ala	Ile	Ser	Ala	Arg	Leu	Gly	Ser																	
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AAA	GTT	AAG	TTG	TCT	TGG	AAG	CTC	TCA	GCT	ATC	ACT	AAG	CTG	GAG	AGC																966	
Lys	Val	Lys	Leu	Ser	Trp	Lys	Leu	Ser	Gly	Ile	Thr	Lys	Leu	Glu	Ser																	
				300				305					310																			
GGA	GGA	TAC	AAC	TTA	ACA	TAT	GAG	ACT	CCA	GAT	GCT	TTA	GTT	TCC	GTG																1014	
Gly	Gly	Tyr	Asn	Leu	Thr	Tyr	Glu	Thr	Pro	Asp	Gly	Leu	Val	Ser	Val																	
			315				320					325																				
CAG	AGC	AAA	AGT	GTT	GTA	ATG	ACG	GTC	CCA	TCT	CAT	GTT	GCA	AGT	GCT																1062	
Gln	Ser	Lys	Ser	Val	Val	Met	Thr	Val	Pro	Ser	His	Val	Ala	Ser	Gly																	
			330			335					340																					
CTC	TTG	COC	CCT	CTT	TCT	GAA	TCT	GCT	GCA	AAT	GCA	CTC	TCA	AAA	CTA																1110	
Leu	Leu	Arg	Pro	Leu	Ser	Glu	Ser	Ala	Ala	Asn	Ala	Leu	Ser	Lys	Leu																	
						350				355					360																	
TAT	TAC	CCA	CCA	GTT	GCA	GCA	GTA	TCT	ATC	TGG	TAC	CCG	AAA	GGA	GCA																1158	
Tyr	Tyr	Pro	Pro	Val	Ala	Ala	Val	Ser	Ile	Ser	Tyr	Pro	Lys	Glu	Ala																	
						365			370				375																			
ATC	GGA	ACA	GAA	TGT	TTG	ATA	GAT	GCT	GAA	CTA	AAG	GCT	TTT	GCG	CAA																1206	
Ile	Arg	Thr	Glu	Cys	Leu	Ile	Asp	Gly	Glu	Leu	Lys	Gly	Phe	Gly	Gln																	
			380					385					390																			
TTG	CAT	CCA	CCG	ACG	CAA	GCA	GTT	GAA	ACA	TTA	GGA	ACT	ATC	TAC	AGC																1254	
Leu	His	Pro	Arg	Thr	Gln	Gly	Val	Glu	Thr	Leu	Gly	Thr	Ile	Tyr	Ser																	
			395				400					405																				
TCC	TCA	CTC	TTT	CCA	AAT	CCG	GCA	CCG	CCG	GGA	AGA	ATT	TTG	CTG	TTG																1302	
Ser	Ser	Leu	Phe	Pro	Asn	Arg	Ala	Pro	Pro	Gly	Arg	Ile	Leu	Leu	Leu																	
			410			415				420																						
AAC	TAC	ATT	GCG	GCG	TCT	ACA	AAC	ACC	GGA	ATT	CTG	TCC	AAG	TCT	GAA																1350	
Asn	Tyr	Ile	Gly	Gly	Ser	Thr	Asn	Thr	Gly	Ile	Leu	Ser	Lys	Ser	Glu																	
						430			435						440																	
GCT	GAG	TTA	GTG	GAA	GCA	GTT	GAC	AGA	GAT	TTG	AGG	AAA	ATG	CTA	ATT																1398	
Gly	Glu	Leu	Val	Glu	Ala	Val	Asp	Arg	Asp	Leu	Arg	Lys	Met	Leu	Ile																	
						445			450					455																		
AAG	CCT	AAT	TGG	ACC	GAT	GCA	CTT	AAA	TTA	GGA	GTT	AGG	GTA	TGG	CCT																1446	

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Lys Pro Asn Ser Thr Asp Pro Leu Lys Leu Gly Val Arg Val Trp Pro
460                               465                               470
CAA GCC ATT CCT CAG TTT CTA GTT GGT CAC TTT GAT ATC CTT GAC ACC 1494
Gln Ala Ile Pro Gln Phe Leu Val Gly His Phe Asp Ile Leu Asp Thr
475                               480                               485
GCT AAA TCA TCT CTA ACG TCT TCG GGC TAC GAA GCG CTA TTT TTG GGT 1542
Ala Lys Ser Ser Leu Thr Ser Ser Gly Tyr Glu Gly Leu Phe Leu Gly
490                               495                               500
GGC AAT TAC GTC GCT GGT GTA GGC TTA GGC CCG TGT GTA GAA GGC GCA 1596
Gly Asn Tyr Val Ala Gly Val Ala Leu Gly Arg Cys Val Glu Gly Ala
505                               510                               515                               520
TAT GAA ACC GCG ATT GAG GTC AAC AAC TTC ATG TCA CCG TAC GCT TAC 1638
Tyr Glu Thr Ala Ile Glu Val Asn Asn Phe Met Ser Arg Tyr Ala Tyr
525                               530                               535
AAG TAAATGTAAA ACATTAAATC TCCAGCTTG CGTGAGTTT ATTAATATTT 1691
Lys

TTGAGATATC CAAAAAATAA AAAAAAA 1719

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(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 537 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

```

Met Glu Leu Ser Leu Leu Arg Pro Thr Thr Gln Ser Leu Leu Pro Ser
1                               5                               10                               15
Phe Ser Lys Pro Asn Leu Arg Leu Asn Val Tyr Lys Pro Leu Arg Leu
20                               25                               30
Arg Cys Ser Val Ala Gly Gly Pro Thr Val Gly Ser Ser Lys Ile Glu
35                               40                               45
Gly Gly Gly Gly Thr Thr Ile Thr Thr Asp Cys Val Ile Val Gly Gly
50                               55                               60
Gly Ile Ser Gly Leu Cys Ile Ala Gln Ala Leu Ala Thr Lys His Pro
65                               70                               75                               80
Asp Ala Ala Pro Asn Leu Ile Val Thr Glu Ala Lys Asp Arg Val Gly
85                               90                               95
Gly Asn Ile Ile Thr Arg Glu Glu Asn Gly Phe Leu Trp Glu Glu Gly
100                               105                               110
Pro Asn Ser Phe Gln Pro Ser Asp Pro Met Leu Thr Met Val Val Asp
115                               120                               125
Ser Gly Leu Lys Asp Asp Leu Val Leu Gly Asp Pro Thr Ala Pro Arg
130                               135                               140

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Phe Val Leu Trp Asn Gly Lys Leu Arg Pro Val Pro Ser Lys Leu Thr
 145 150 155 160
 Asp Leu Pro Phe Phe Asp Leu Met Ser Ile Gly Gly Lys Ile Arg Ala
 165 170 175
 Gly Phe Gly Ala Leu Gly Ile Arg Pro Ser Pro Pro Gly Arg Glu Glu
 180 185 190
 Ser Val Glu Glu Phe Val Arg Arg Asn Leu Gly Asp Glu Val Phe Glu
 195 200 205
 Arg Leu Ile Glu Pro Phe Cys Ser Gly Val Tyr Ala Gly Asp Pro Ser
 210 215 220
 Lys Leu Ser Met Lys Ala Ala Phe Gly Lys Val Trp Lys Leu Glu Glu
 225 230 235 240
 Asn Gly Gly Ser Ile Ile Gly Gly Thr Phe Lys Ala Ile Glu Glu Arg
 245 250 255
 Lys Asn Ala Pro Lys Ala Glu Arg Asp Pro Arg Leu Pro Lys Pro Glu
 260 265 270
 Gly Glu Thr Val Gly Ser Phe Arg Lys Gly Leu Arg Met Leu Pro Glu
 275 280 285
 Ala Ile Ser Ala Arg Leu Gly Ser Lys Val Lys Leu Ser Trp Lys Leu
 290 295 300
 Ser Gly Ile Thr Lys Leu Glu Ser Gly Gly Tyr Asn Leu Thr Tyr Glu
 305 310 315 320
 Thr Pro Asp Gly Leu Val Ser Val Glu Ser Lys Ser Val Val Met Thr
 325 330 335
 Val Pro Ser His Val Ala Ser Gly Leu Leu Arg Pro Leu Ser Glu Ser
 340 345 350
 Ala Ala Asn Ala Leu Ser Lys Leu Tyr Tyr Pro Pro Val Ala Ala Val
 355 360 365
 Ser Ile Ser Tyr Pro Lys Glu Ala Ile Arg Thr Glu Cys Leu Ile Asp
 370 375 380
 Gly Glu Leu Lys Gly Phe Gly Glu Leu His Pro Arg Thr Glu Gly Val
 385 390 395 400
 Glu Thr Leu Gly Thr Ile Tyr Ser Ser Ser Leu Phe Pro Asn Arg Ala
 405 410 415
 Pro Pro Gly Arg Ile Leu Leu Leu Asn Tyr Ile Gly Gly Ser Thr Asn
 420 425 430
 Thr Gly Ile Leu Ser Lys Ser Glu Gly Glu Leu Val Glu Ala Val Asp
 435 440 445
 Arg Asp Leu Arg Lys Met Leu Ile Lys Pro Asn Ser Thr Asp Pro Leu
 450 455 460
 Lys Leu Gly Val Arg Val Trp Pro Glu Ala Ile Pro Glu Phe Leu Val
 465 470 475 480

Gly His Phe Asp Ile Leu Asp Thr Ala Lys Ser Ser Leu Thr Ser Ser
483 490 495
Gly Tyr Glu Gly Leu Phe Leu Gly Gly Asn Tyr Val Ala Gly Val Ala
500 505 510
Leu Gly Arg Cys Val Glu Gly Ala Tyr Glu Thr Ala Ile Glu Val Asn
515 520 525
Asn Phe Met Ser Arg Tyr Ala Tyr Lys
530 535

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1738 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 70..1596
- (D) OTHER INFORMATION: /note= 'Arabidopsis protein-2 cDNA; sequence from pADC-1'

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

TTTCTTACTT ATTCCGCTCA CTGCTTCCA CTGCTCAGAG ATTTCGACTC TGAATTCTG	60
CAGATAOCA ATG GCG TCT GGA GCA GTA GCA GAT CAT CAA ATT GAA GCG	108
Met Ala Ser Gly Ala Val Ala Asp His Gln Ile Glu Ala	
1 5 10	
GTT TCA GGA AAA AGA GTC GCA GTC GTA GGT GCA GGT GTA AGT GGA CTT	156
Val Ser Gly Lys Arg Val Ala Val Val Gly Ala Gly Val Ser Gly Leu	
15 20 25	
GCG GCG GCT TAC AAG TTG AAA TCG AGG GGT TTG AAT GTC ACT GTG TTT	204
Ala Ala Ala Tyr Lys Leu Lys Ser Arg Gly Leu Asn Val Thr Val Phe	
30 35 40 45	
GAA GCT GAT GGA AGA GTA GGT GCG AAG TTG AGA AGT GTT ATG CAA AAT	252
Glu Ala Asp Gly Arg Val Gly Gly Lys Leu Arg Ser Val Met Gln Asn	
50 55 60	
GGT TTG ATT TGG GAT GAA GGA GCA AAC ACC ATG ACT GAG GCT GAG CCA	300
Gly Leu Ile Trp Asp Glu Gly Ala Asn Thr Met Thr Glu Ala Glu Pro	
65 70 75	
GAA GTT GCG AGT TTA CTT GAT GAT CTT GCG CTT COT GAG AAA CAA CAA	348
Glu Val Gly Ser Leu Leu Asp Asp Leu Gly Leu Arg Glu Lys Gln Gln	
80 85 90	

TTT	CCA	ATT	TCA	CAG	AAA	AAG	CGG	TAT	ATT	GTC	CGG	AGT	GCT	GTA	CCT	396
Phe	Pro	Ile	Ser	Gln	Lys	Lys	Arg	Tyr	Ile	Val	Arg	Asn	Gly	Val	Pro	
95					100					105						
GTC	ATG	CTA	CCT	ACC	AAT	CCC	ATA	GAG	CTG	GTC	ACA	AGT	AGT	GTC	CTC	444
Val	Met	Leu	Pro	Thr	Asn	Pro	Ile	Glu	Leu	Val	Thr	Ser	Ser	Val	Leu	
110					115					120					125	
TCT	ACC	CAA	TCT	AAG	TTT	CAA	ATC	TTG	TTG	GAA	CCA	TTT	TTA	TGG	AAG	492
Ser	Thr	Gln	Ser	Lys	Phe	Gln	Ile	Leu	Leu	Glu	Pro	Phe	Leu	Tyr	Lys	
				130					135					140		
AAA	AAG	TCC	TCA	AAA	GTC	TCA	GAT	GCA	TCT	GCT	GAA	GAA	AGT	GTA	ACC	540
Lys	Lys	Ser	Ser	Lys	Val	Ser	Asp	Ala	Ser	Ala	Glu	Glu	Ser	Val	Ser	
				145				150					155			
GAG	TTC	TTT	CAA	CGC	CAT	TTT	GGA	CAA	GAG	GTT	GTT	GAC	TAT	CTC	ATC	588
Glu	Phe	Phe	Gln	Arg	His	Phe	Gly	Gln	Glu	Val	Val	Asp	Tyr	Leu	Ile	
		160					165					170				
GAC	CCT	TTT	GTT	GCT	GGA	ACA	AGT	GCT	GCG	GAC	CCT	GAT	TCC	CCT	TCA	636
Asp	Pro	Phe	Val	Gly	Gly	Thr	Ser	Ala	Ala	Asp	Pro	Asp	Ser	Leu	Ser	
		175				180					185					
ATG	AAG	CAT	TCT	TTC	CCA	GAT	CTC	TGG	AAT	GTA	GAG	AAA	AGT	TTT	GCC	684
Met	Lys	His	Ser	Phe	Pro	Asp	Leu	Tyr	Asn	Val	Glu	Lys	Ser	Phe	Gly	
					195					200					205	
TCT	ATT	ATA	GTC	GCT	GCA	ATC	AGA	ACA	AAG	TTT	GCT	GCT	AAA	GCT	GCT	732
Ser	Ile	Ile	Val	Gly	Ala	Ile	Arg	Thr	Lys	Phe	Ala	Ala	Lys	Gly	Gly	
				210					215					220		
AAA	AGT	AGA	GAC	ACA	AAG	AGT	TCT	CCT	GCC	ACA	AAA	AAG	GCT	TCC	CCT	780
Lys	Ser	Arg	Asp	Thr	Lys	Ser	Ser	Pro	Gly	Thr	Lys	Lys	Gly	Ser	Arg	
			225					230					235			
GCG	TCA	TTC	TCT	TTT	AAG	GCG	GGA	ATG	CAG	ATT	CTT	CCT	GAT	ACC	TTG	828
Gly	Ser	Phe	Ser	Phe	Lys	Gly	Met	Gln	Ile	Leu	Pro	Asp	Thr	Leu		
			240				245					250				
TGC	AAA	AGT	CTC	TCA	CAT	GAT	GAG	ATC	AAT	TTA	GAC	TCC	AAG	GTA	CTC	876
Cys	Lys	Ser	Leu	Ser	His	Asp	Glu	Ile	Asn	Leu	Asp	Ser	Lys	Val	Leu	
			255			260					265					
TCT	TTG	TCT	TAC	AAT	TCT	GGA	TCA	AGA	CAG	GAG	AAC	TGG	TCA	TTA	TCT	924
Ser	Leu	Ser	Tyr	Asn	Ser	Gly	Ser	Arg	Gln	Asn	Tyr	Ser	Leu	Ser		
					275					280				285		
TGT	GTT	TCG	CAT	AAT	GAA	ACG	CAG	AGA	CAA	AAC	CCC	CAT	TAT	GAT	GCT	972
Cys	Val	Ser	His	Asn	Glu	Thr	Gln	Arg	Gln	Asn	Pro	His	Tyr	Asp	Ala	
					290				295					300		
GTA	ATT	ATG	ACG	GCT	CCT	CTG	TGC	AAT	GTC	AAG	GAG	ATG	AAG	GTT	ATG	1020
Val	Ile	Met	Thr	Ala	Pro	Leu	Cys	Asn	Val	Lys	Glu	Met	Lys	Val	Met	
					305			310					315			
AAA	GGA	GGA	CAA	CCC	TTT	CAG	CTA	AAC	TTT	CTC	CCC	GAG	ATT	AAT	TAC	1068
Lys	Gly	Gly	Gln	Pro	Phe	Gln	Leu	Asn	Phe	Leu	Pro	Glu	Ile	Asn	Tyr	
					320			325				330				
ATG	CCC	CTC	TCG	GTT	TTA	ATC	ACC	ACA	TTC	ACA	AAG	GAG	AAA	GTA	AAG	1116
Met	Pro	Leu	Ser	Val	Leu	Ile	Thr	Thr	Phe	Thr	Lys	Glu	Lys	Val	Lys	

335	AGA CCT CTT GAA GGC TTT GGG GTA CTC ATT CCA TCT AAG GAG CAA AAG	1164
Arg Pro Leu Glu Gly Phe Gly Val Leu Ile Pro Ser Lys Glu Glu Lys	350 355 360 365	
CAT GGT TTC AAA ACT CTA GGT ACA CTT TTT TCA TCA ATG ATG TTT CCA	1212	
His Gly Phe Lys Thr Leu Gly Thr Leu Phe Ser Ser Met Met Phe Pro	370 375 380	
GAT GGT TCC CCT AGT GAC GTT CAT CTA TAT ACA ACT TTT ATT GGT GGG	1260	
Asp Arg Ser Pro Ser Asp Val His Leu Tyr Thr Thr Phe Ile Gly Gly	385 390 395	
AGT AAG AAC CAG GAA CTA GCC AAA GCT TCC ACT GAC GAA TTA AAA CAA	1300	
Ser Arg Asn Gln Glu Leu Ala Lys Ala Ser Thr Asp Glu Leu Lys Glu	400 405 410	
GTT GTG ACT TCT GAC CTT CAG CGA CTG TTG GGG GTT GAA GGT GAA CCC	1356	
Val Val Thr Ser Asp Leu Gln Arg Leu Leu Gly Val Glu Gly Glu Pro	415 420 425	
GTG TCT GTC AAC CAT TAC TAT TGG AGG AAA GCA TTC CCG TTG TAT GAC	1404	
Val Ser Val Asn His Tyr Tyr Trp Arg Lys Ala Phe Pro Leu Tyr Asp	430 435 440 445	
AGC AOC TAT GAC TCA GTC ATG GAA GCA ATT GAC AAG ATG GAG AAT GAT	1452	
Ser Ser Tyr Asp Ser Val Met Glu Ala Ile Asp Lys Met Glu Asn Asp	450 455 460	
CTA CCT GGG TTC TTC TAT CCA GGT AAT CAT CGA GGG GGG CTC TCT GTT	1500	
Leu Pro Gly Phe Tyr Ala Gly Asn His Arg Gly Gly Leu Ser Val	465 470 475	
GGG AAA TCA ATA GCA TCA GGT TGC AAA GCA GCT GAC CTT GTG ATC TCA	1540	
Gly Lys Ser Ile Ala Ser Gly Cys Lys Ala Ala Asp Leu Val Ile Ser	480 485 490	
TAC CTG GAG TCT TGC TCA AAT GAC AAG AAA CCA AAT GAC AGC TTA TAACTTCTC	1603	
Tyr Leu Glu Ser Cys Ser Asn Asp Lys Lys Pro Asn Asp Ser Leu	495 500 505	
AAAGTTCTCTC CCTTTTATC ACTTACTTTG TAACTTCTA AAATGCAACA AGCCCCCTG	1663	
CGATTAGCCA ACAAATGAGC AAAGCCGGA TTCTCATAG GCTCCTAAT TCCAGATTA	1723	
ACTATTTATG TAAAA	1738	

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 508 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ala Ser Gly Ala Val Ala Asp His Gln Ile Glu Ala Val Ser Gly
 1 5 10 15

Lys Arg Val Ala Val Val Gly Ala Gly Val Ser Gly Leu Ala Ala Ala
 20 25 30
 Tyr Lys Leu Lys Ser Arg Gly Leu Asn Val Thr Val Phe Glu Ala Asn
 35 40 45
 Gly Arg Val Gly Gly Lys Leu Arg Ser Val Met Gln Asn Gly Leu Ile
 50 55 60
 Trp Asp Glu Gly Ala Asn Thr Met Thr Glu Ala Glu Pro Glu Val Gly
 65 70 75 80
 Ser Leu Leu Asp Asp Leu Gly Leu Arg Glu Lys Gln Gln Phe Pro Ile
 85 90 95
 Ser Gln Lys Lys Arg Tyr Ile Val Arg Asn Gly Val Pro Val Met Leu
 100 105 110
 Pro Thr Asn Pro Ile Glu Leu Val Thr Ser Ser Val Leu Ser Thr Gln
 115 120 125
 Ser Lys Phe Gln Ile Leu Leu Glu Pro Phe Leu Trp Lys Lys Lys Ser
 130 135 140
 Ser Lys Val Ser Asp Ala Ser Ala Glu Glu Ser Val Ser Glu Phe Phe
 145 150 155 160
 Gln Arg His Phe Gly Gln Glu Val Val Asp Tyr Leu Ile Asp Pro Phe
 165 170 175
 Val Gly Gly Thr Ser Ala Ala Asn Pro Asp Ser Leu Ser Met Lys His
 180 185 190
 Ser Phe Pro Asp Leu Trp Asn Val Glu Lys Ser Phe Gly Ser Ile Ile
 195 200 205
 Val Gly Ala Ile Arg Thr Lys Phe Ala Ala Lys Gly Gly Lys Ser Arg
 210 215 220
 Asp Thr Lys Ser Ser Pro Gly Thr Lys Lys Gly Ser Arg Gly Ser Phe
 225 230 235 240
 Ser Phe Lys Gly Gly Met Gln Ile Leu Pro Asp Thr Leu Cys Lys Ser
 245 250 255
 Leu Ser His Asp Glu Ile Asn Leu Asp Ser Lys Val Leu Ser Leu Ser
 260 265 270
 Tyr Asn Ser Gly Ser Arg Gln Glu Asn Trp Ser Leu Ser Cys Val Ser
 275 280 285
 His Asn Glu Thr Gln Arg Gln Asn Pro His Tyr Asp Ala Val Ile Met
 290 295 300
 Thr Ala Pro Leu Cys Asn Val Lys Glu Met Lys Val Met Lys Gly Gly
 305 310 315 320
 Gln Pro Phe Gln Leu Asn Phe Leu Pro Glu Ile Asn Tyr Met Pro Leu
 325 330 335
 Ser Val Leu Ile Thr Thr Phe Thr Lys Glu Lys Val Lys Arg Pro Leu
 340 345 350

Glu Gly Phe Gly Val Leu Ile Pro Ser Lys Glu Gln Lys His Gly Phe
355 360 365
Lys Thr Leu Gly Thr Leu Phe Ser Ser Met Met Phe Pro Asp Arg Ser
370 375 380
Pro Ser Asp Val His Leu Tyr Thr Thr Phe Ile Gly Gly Ser Arg Asn
385 390 395 400
Gln Glu Leu Ala Lys Ala Ser Thr Asp Glu Leu Lys Gln Val Val Thr
405 410 415
Ser Asp Leu Gln Arg Leu Leu Gly Val Glu Gly Glu Pro Val Ser Val
420 425 430
Asn His Tyr Tyr Trp Arg Lys Ala Phe Pro Leu Tyr Asp Ser Ser Tyr
435 440 445
Asp Ser Val Met Glu Ala Ile Asp Lys Met Glu Asn Asp Leu Pro Gly
450 455 460
Phe Phe Tyr Ala Gly Asn His Arg Gly Gly Leu Ser Val Gly Lys Ser
465 470 475 480
Ile Ala Ser Gly Cys Lys Ala Ala Asp Leu Val Ile Ser Tyr Leu Glu
485 490 495
Ser Cys Ser Asn Asp Lys Lys Pro Asn Asp Ser Leu
500 505

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1698 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 2..1453
- (D) OTHER INFORMATION: /note= "Maize proteox-1 cDNA (not full-length); sequence from pMDC-4"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

G	AAT	TGG	CCG	GAC	TGC	GTC	GTC	GTC	GCC	GGA	GCC	ATC	AGT	GCC	CTC	46
Asn	Ser	Ala	Asp	Cys	Val	Val	Val	Gly	Gly	Gly	Ile	Ser	Gly	Leu		
1				5				10					15			
TGC	ACC	GCG	CAG	GCG	CTG	GCC	ACG	CGG	CAC	GCC	GTC	GCG	GAC	GTC	CTT	94
Cys	Thr	Ala	Gln	Ala	Leu	Ala	Thr	Arg	His	Gly	Val	Gly	Asp	Val	Leu	
				20				25					30			

GTC	ACG	GAG	GCC	GCC	GCC	GCC	GCC	GCC	GCC	ACC	ATT	ACC	ACC	GTC	GAG	162
Val	Thr	Glu	Ala	Arg	Ala	Arg	Pro	Gly	Gly	Asn	Ile	Thr	Thr	Val	Glu	
			35					40					45			
GCC	GCC	GAG	GAA	GCG	TAC	CTC	TGG	GAG	GAG	GGT	CCC	AAC	ACC	TTC	CAG	190
Arg	Pro	Glu	Glu	Gly	Tyr	Leu	Trp	Glu	Glu	Gly	Pro	Asn	Ser	Phe	Gln	
		50					55					60				
CCC	TCC	GAC	CCC	GTT	CTC	ACC	ATG	GCC	GTC	GAC	AGC	GGA	CTG	AAG	GAT	230
Pro	Ser	Asp	Pro	Val	Leu	Thr	Met	Ala	Val	Asp	Ser	Gly	Leu	Lys	Asp	
		65				70					75					
GAC	TTC	GTT	TTT	GCG	GAC	CCA	AAC	GCG	CCG	GCT	TTC	GTC	CTG	TGG	GAG	286
Asp	Leu	Val	Phe	Gly	Asp	Pro	Asn	Ala	Pro	Arg	Phe	Val	Leu	Trp	Glu	
		80			85					90					95	
GCG	AAG	CTG	AGG	CCC	GTC	CCA	TCC	AAG	CCC	GCC	GAC	CTC	CCG	TTC	TTC	334
Gly	Lys	Leu	Arg	Pro	Val	Pro	Ser	Lys	Pro	Ala	Asp	Leu	Pro	Phe	Phe	
			100						105					110		
GAT	CTC	ATG	AGC	ATC	CCA	GCG	AAG	CTC	AGG	GCC	GCT	CTA	GAC	GCG	CTT	382
Asp	Leu	Met	Ser	Ile	Pro	Gly	Lys	Leu	Arg	Ala	Gly	Leu	Gly	Ala	Leu	
			115					120					125			
GCC	ATC	GCC	CCG	GCT	GCT	CCA	GCC	GCG	GAA	GAG	TCA	GTC	GAG	GAG	TTC	430
Gly	Ile	Arg	Pro	Pro	Pro	Pro	Gly	Arg	Glu	Glu	Ser	Val	Glu	Glu	Phe	
		130					135					140				
GTC	GCC	CCG	AAC	CTC	GCT	GCT	GAG	GTC	TTT	GAG	CCC	CTC	ATT	GAG	GCT	478
Val	Arg	Arg	Asn	Leu	Gly	Ala	Glu	Val	Phe	Glu	Arg	Leu	Ile	Glu	Pro	
		145				150					155					
TTC	TGC	TCA	GCT	GTC	TAT	GCT	GCT	GAT	CTT	TCT	AAG	CTC	ACC	ATG	AAG	526
Phe	Cys	Ser	Gly	Val	Tyr	Ala	Gly	Asp	Pro	Ser	Lys	Leu	Ser	Met	Lys	
		160			165				170						175	
GCT	GCA	TTT	GCG	AAG	GTT	TGG	CCG	TTC	GAA	GAA	ACT	GGA	GCT	AGT	ATT	574
Ala	Ala	Phe	Gly	Lys	Val	Trp	Arg	Leu	Glu	Glu	Thr	Gly	Gly	Ser	Ile	
			180					185						190		
ATT	GCT	GGA	ACC	ATC	AAG	ACA	ATT	CAG	GAG	AGG	AGC	AAG	AAT	CCA	AAA	622
Ile	Gly	Gly	Thr	Ile	Lys	Thr	Ile	Gln	Glu	Arg	Ser	Lys	Asn	Pro	Lys	
			195				200						205			
CCA	CCG	AGG	GAT	GCC	GCG	CTT	CCG	AAG	CCA	AAA	GCG	CAG	ACA	GTT	GCA	670
Pro	Pro	Arg	Asp	Ala	Arg	Leu	Pro	Lys	Pro	Lys	Gly	Gln	Thr	Val	Ala	
		210					215					220				
TCT	TTC	AGG	AAG	GCT	CTT	GCC	ATG	CTT	CCA	AAT	GCC	ATT	ACA	TCC	AGC	718
Ser	Phe	Arg	Lys	Gly	Leu	Ala	Met	Leu	Pro	Asn	Ala	Ile	Thr	Ser	Ser	
		225				230					235					
TTC	GCT	AGT	AAA	GTC	AAA	CTA	TCA	TGG	AAA	CTC	ACG	AGC	ATT	ACA	AAA	766
Leu	Gly	Ser	Lys	Val	Lys	Leu	Ser	Trp	Lys	Leu	Thr	Ser	Ile	Thr	Lys	
		240			245					250					255	
TCA	GAT	GAC	AAG	GGA	TAT	GTT	TTC	GAG	TAT	GAA	ACG	CCA	GAA	GCG	GTT	814
Ser	Asp	Asp	Lys	Gly	Tyr	Val	Leu	Glu	Tyr	Glu	Thr	Pro	Glu	Gly	Val	
				260				265						270		
GTT	TGG	GTC	CAG	GCT	AAA	AGT	GTT	ATC	ATG	ACT	ATT	CCA	TCA	TAT	GTT	862
Val	Ser	Val	Gln	Ala	Lys	Ser	Val	Ile	Met	Thr	Ile	Pro	Ser	Tyr	Val	
			275				280						285			

GCT AGC AAC ATT TTG CGT CCA CTT TCA AGC GAT GCT GCA GAT GCT CTA Ala Ser Asn Ile Leu Arg Pro Leu Ser Ser Asp Ala Ala Asp Ala Leu 290 295 300	910
TCA AGA TTC TAT TAT CCA CCG GTT OCT GCT GTA ACT GTT TCG TAT CCA Ser Arg Phe Tyr Tyr Pro Pro Val Ala Ala Val Thr Val Ser Tyr Pro 305 310 315	950
AGG GAA GCA ATT AGA AAA GAA TGC TTA ATT GAT GCG GAA CTC CAG GGC Lys Glu Ala Ile Arg Lys Glu Cys Leu Ile Asp Gly Glu Leu Gln Gly 320 325 330 335	1006
TTT GGC CAG TTG CAT CCA CGT AGT CAA GGA GTT GAG ACA TTA GGA ACA Phe Gly Gln Leu His Pro Arg Ser Gln Gly Val Glu Thr Leu Gly Thr 340 345 350	1034
ATA TAC AGT TCC TCA CTC TTT CCA AAT COT GCT CTT GAC GGT AGC GTG Ile Tyr Ser Ser Ser Leu Phe Pro Asn Arg Ala Pro Asp Gly Arg Val 355 360 365	1102
TTA CTT CTA AAC TAC ATA GGA GGT GCT ACA AAC ACA GGA ATT GTT TCC Leu Leu Leu Asn Tyr Ile Gly Gly Ala Thr Asn Thr Gly Ile Val Ser 370 375 380	1150
AGG ACT GAA AGT GAG CTG GTC GAA GCA GTT GAC COT GAC CTC GGA AAA Lys Thr Glu Ser Glu Leu Val Glu Ala Val Asp Arg Asp Leu Arg Lys 385 390 395	1190
ATG CTT ATA AAT TCT ACA GCA GTG GAC COT TTA GTC CTT GGT GTT GGA Met Leu Ile Asn Ser Thr Ala Val Asp Pro Leu Val Leu Gly Val Arg 400 405 410 415	1246
GTT TCG CCA CAA GCC ATA CCT CAG TTC CTG GTA GGA CAT CTT GAT CTT Val Trp Pro Gln Ala Ile Pro Gln Phe Leu Val Gly His Leu Asp Leu 420 425 430	1294
CTG GAA GCC GCA AAA GCT GCC CTG GAC GGA GGT GGC TAC GAT GCG CTG Leu Glu Ala Ala Lys Ala Ala Leu Asp Arg Gly Gly Tyr Asp Gly Leu 435 440 445	1342
TTC CTA GGA GCG AAC TAT GTT GCA GGA GTT GCC CTG GGC AGA TGC GTT Phe Leu Gly Gly Asn Tyr Val Ala Gly Val Ala Leu Gly Arg Cys Val 450 455 460	1390
GAG GGC GCG TAT GAA AGT GCC TCG CAA ATA TCT GAC TTC TTG ACC AAG Glu Gly Ala Tyr Glu Ser Ala Ser Gln Ile Ser Asp Phe Leu Thr Lys 465 470 475	1438
TAT GCC TAC AAG TGATGAAGA AOTGAGGCC TACTGTGTA TCGTTTATGT Tyr Ala Tyr Lys 480	1490
TGCATAGATG AGGTGCTCC GGGGAAAAA AAGCTTGAAT AGTATTTTTT ATTCTTATTT	1550
TGTAAATTC ATTCTGTTT TTTTCTAT CAGTAATTAG TTATATTTTA GTTCTGTAGG	1610
AGATTGTTCT GTTCACTGCC CTTCAAAAGA AATTTTATTT TTCAATCTTT TATGAGAGCT	1670
GTCTACTTA AAAAAAAA AAAAAAA	1698

(2) INFORMATION FOR SEQ ID NO:6:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 403 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

```

Asn Ser Ala Asp Cys Val Val Val Gly Gly Gly Ile Ser Gly Leu Cys
 1          5          10          15
Thr Ala Gln Ala Leu Ala Thr Arg His Gly Val Gly Asp Val Leu Val
 20          25          30
Thr Glu Ala Arg Ala Arg Pro Gly Gly Asn Ile Thr Thr Val Glu Arg
 35          40          45
Pro Glu Glu Gly Tyr Leu Trp Gln Gln Gly Pro Asn Ser Phe Gln Pro
 50          55          60
Ser Asp Pro Val Leu Thr Met Ala Val Asp Ser Gly Leu Lys Asp Asp
 65          70          75          80
Leu Val Phe Gly Asp Pro Asn Ala Pro Arg Phe Val Leu Trp Gln Gly
 85          90          95
Lys Leu Arg Pro Val Pro Ser Lys Pro Ala Asp Leu Pro Phe Phe Asp
100          105          110
Leu Met Ser Ile Pro Gly Lys Leu Arg Ala Gly Leu Gly Ala Leu Gly
115          120          125
Ile Arg Pro Pro Pro Pro Gly Arg Gln Glu Ser Val Glu Glu Phe Val
130          135          140
Arg Arg Asn Leu Gly Ala Glu Val Phe Glu Arg Leu Ile Glu Pro Phe
145          150          155          160
Cys Ser Gly Val Tyr Ala Gly Asp Pro Ser Lys Leu Ser Met Lys Ala
165          170          175
Ala Phe Gly Lys Val Trp Arg Leu Glu Glu Thr Gly Gly Ser Ile Ile
180          185          190
Gly Gly Thr Ile Lys Thr Ile Gln Glu Arg Ser Lys Asn Pro Lys Pro
195          200          205
Pro Arg Asp Ala Arg Leu Pro Lys Pro Lys Gly Gln Thr Val Ala Ser
210          215          220
Phe Arg Lys Gly Leu Ala Met Leu Pro Asn Ala Ile Thr Ser Ser Leu
225          230          235          240
Gly Ser Lys Val Lys Leu Ser Trp Lys Leu Thr Ser Ile Thr Lys Ser
245          250          255
Asp Asp Lys Gly Tyr Val Leu Glu Tyr Glu Thr Pro Glu Gly Val Val
260          265          270
Ser Val Gln Ala Lys Ser Val Ile Met Thr Ile Pro Ser Tyr Val Ala
275          280          285

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Ser Asn Ile Leu Arg Pro Leu Ser Ser Asp Ala Ala Asp Ala Leu Ser
290                295                300

Arg Phe Tyr Tyr Pro Pro Val Ala Ala Val Thr Val Ser Tyr Pro Lys
305                310                315                320

Glu Ala Ile Arg Lys Glu Cys Leu Ile Asp Gly Glu Leu Glu Gly Phe
325                330                335

Gly Gln Leu His Pro Arg Ser Gln Gly Val Glu Thr Leu Gly Thr Ile
340                345                350

Tyr Ser Ser Ser Leu Phe Pro Asn Arg Ala Pro Asp Gly Arg Val Leu
355                360                365

Leu Leu Asn Tyr Ile Gly Gly Ala Thr Asn Thr Gly Ile Val Ser Lys
370                375                380

Thr Glu Ser Glu Leu Val Glu Ala Val Asp Arg Asp Leu Arg Lys Ser
385                390                395                400

Leu Ile Asn Ser Thr Ala Val Asp Pro Leu Val Leu Gly Val Arg Val
405                410                415

Trp Pro Gln Ala Ile Pro Gln Phe Leu Val Gly His Leu Asp Leu Leu
420                425                430

Glu Ala Ala Lys Ala Ala Leu Asp Arg Gly Gly Tyr Asp Gly Leu Phe
435                440                445

Leu Gly Gly Asn Tyr Val Ala Gly Val Ala Leu Gly Arg Cys Val Glu
450                455                460

Gly Ala Tyr Glu Ser Ala Ser Gln Ile Ser Asp Phe Leu Thr Lys Tyr
465                470                475                480

Ala Tyr Lys

```

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2061 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 64..1698
- (D) OTHER INFORMATION: /note= "Maize protein-2 cDNA; sequence from pMDC-3"

(21) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CTCTCTAC	TCACCTCCA	GCACACACAG	CAATCTCCA	TCCTCTCCA	AGCCTACT	60
CAA ATG CTC GCT TGG ACT GCG TCA GCG TCA TCC GCT TCG TCC CAG CCG	100					
Met Leu Ala Leu Thr Ala Ser Ala Ser Ser Ala Ser Ser His Pro	15					
TAT GCG CAC GCG TCC GCG CAC ACT GGT GCG CCG CCG CTA GGT GCG GTC	156					
Tyr Arg His Ala Ser Ala His Thr Arg Arg Pro Arg Leu Arg Ala Val	20					
CTC GCG ATG GCG GCG TCC GAC GAC GCG GGT GCA GCG CCG GCG AAG TCG	204					
Leu Ala Met Ala Gly Ser Asp Asp Pro Arg Ala Ala Pro Ala Arg Ser	35					
GTC GCG GTC GTC GCG GCG GCG GTC AAG GCG CTC GCG GCG GCG TAC AAG	252					
Val Ala Val Val Gly Ala Gly Val Ser Gly Leu Ala Ala Ala Tyr Arg	50					
CTC AAG CAG ACG GCG GTC AAG GTC ACG GTC TTC GAA GCG GCG GAC AAG	300					
Leu Arg Gln Ser Gly Val Asn Val Thr Val Phe Gly Ala Ala Asp Arg	65					
GCG GGA GGA AAG ATA GCG ACC AAT TCC GAG GCG GCG TTT GTC TCG GAT	348					
Ala Gly Gly Lys Ile Arg Thr Asn Ser Glu Gly Gly Phe Val Trp Asp	80					
GAA GAA GCT AAG ACC ATG ACA GAA GGT GAA TCG GAG GCG AAT AAG CTG	396					
Glu Gly Ala Asn Thr Met Thr Glu Gly Glu Trp Glu Ala Ser Arg Leu	100					
ATT GAT GAT CTT GGT CTA CAA GAC AAA CAG CAG TAT CCT AAG TCC CAA	444					
Ile Asp Asp Cys Gly Leu Gln Asp Lys Gln Gln Tyr Pro Asn Ser Gln	115					
CAC AAG GGT TAC ATT GTC AAA GAT GGA GCA CCA CCA CTG ATT CCG TCG	492					
His Lys Arg Tyr Ile Val Lys Asp Gly Ala Pro Ala Leu Ile Pro Ser	130					
GAT CCC ATT TCG CTA ATG AAA AGC AGT GTT CTT TCG ACA AAA TCA AAG	540					
Asp Pro Ile Ser Leu Met Lys Ser Ser Val Leu Ser Thr Lys Ser Lys	145					
ATT GCG TTA TTT TTT GAA CCA TTT CTC TAC AAG AAA GCT AAG ACA AGA	588					
Ile Ala Leu Phe Phe Glu Pro Phe Leu Tyr Lys Lys Ala Asn Thr Arg	160					
AAC TCT GGA AAA GTG TCT GAG GAG CAC TTG AGT GAG AGT GTT GCG AGC	636					
Asn Ser Gly Lys Val Ser Glu Glu His Leu Ser Glu Ser Val Gly Ser	180					
TTC TGT GAA CCG CAC TTT GGA AGA GAA GTT GTT GAC TAT TTT GTT GAT	684					
Phe Cys Glu Arg His Phe Gly Arg Glu Val Val Asp Tyr Phe Val Asp	195					
CCA TTT GTA GGT GGA ACA AGT GCA GGA GAT CCA GAG TCA CTA TCT ATT	732					
Pro Phe Val Ala Gly Thr Ser Ala Gly Asp Pro Glu Ser Leu Ser Ile	210					
GGT CAT GCA TTC CCA GCA TTG TGG AAT TTG GAA AGA AAG TAT GGT TCA	780					
Arg His Ala Phe Pro Ala Leu Trp Asn Leu Glu Arg Lys Tyr Gly Ser	225					

GTT	ATT	GTT	GCT	GCC	ATC	TTC	TCT	AAG	CYA	GCA	GCT	AAA	GCT	GCT	GCA	828
Val	Ile	Val	Gly	Ala	Ile	Leu	Ser	Lys	Leu	Ala	Ala	Lys	Gly	Asp	Pro	
240				245					250						255	
GTA	AAG	ACA	AGA	CAT	GAT	TCA	TCA	GCG	AAA	AGA	AGG	AAT	AGA	GGA	GTG	876
Val	Lys	Thr	Arg	His	Asp	Ser	Ser	Gly	Lys	Arg	Arg	Asn	Arg	Arg	Val	
				260				265							270	
TCC	TTT	TCA	TTT	CAT	GCT	GGA	ATG	CAG	TCA	CYA	ATA	AAT	GCA	CTT	CAC	924
Ser	Phe	Ser	Phe	His	Gly	Gly	Met	Gln	Ser	Leu	Ile	Asn	Ala	Leu	His	
			275					280					285			
AAT	GAA	GTT	GGA	GAT	GAT	AAT	GTG	AAG	CTT	GCT	ACA	GAA	GTG	TTG	TCA	972
Asn	Glu	Val	Gly	Asp	Asp	Asn	Val	Lys	Leu	Gly	Thr	Glu	Val	Leu	Ser	
		290					295					300				
TTG	GCA	TGT	ACA	TTT	GAT	GGA	GTT	CCT	GCA	CYA	GSC	AGG	TGG	TCA	ATT	1020
Leu	Ala	Cys	Thr	Phe	Asp	Gly	Val	Pro	Ala	Leu	Gly	Arg	Trp	Ser	Ile	
						310						315				
TCT	GTT	GAT	TGG	AAG	GAT	AGC	GCT	GAC	AAG	GAC	CTT	GCT	AGT	AAC	CAA	1068
Ser	Val	Asp	Ser	Lys	Asp	Ser	Gly	Asp	Lys		Leu	Ala	Ser	Asn	Gln	
					325				330						335	
ACC	TTT	GAT	GCT	GTT	ATA	ATG	ACA	GCT	CCA	TTG	TCA	AAT	GTC	GCG	AGG	1116
Thr	Phe	Asp	Ala	Val	Ile	Met	Thr	Ala	Pro	Leu	Ser	Asn	Val	Arg	Arg	
					340				345					350		
ATG	AAG	TTC	ACC	AAA	GCT	GGA	GCT	CCG	GTT	GTT	CTT	GAC	TTT	CTT	CCT	1164
Met	Lys	Phe	Thr	Lys	Gly	Gly	Ala	Pro	Val	Val	Leu	Asp	Phe	Leu	Pro	
			355					360					365			
AAG	ATG	GAT	TAT	CTA	CCA	CTA	TCT	CTC	ATG	GTG	ACT	GCT	TTT	AAG	AAG	1212
Lys	Met	Asp	Tyr	Leu	Pro	Leu	Ser	Leu	Met	Val	Thr	Ala	Phe	Lys	Lys	
			370				375					380				
GAT	GAT	GTC	AAG	AAA	CCT	CTG	GAA	GGA	TTT	GCG	GTC	TTA	ATA	CCT	TAC	1260
Asp	Asp	Val	Lys	Lys	Pro	Leu	Glu	Gly	Phe	Gly	Val	Leu	Ile	Pro	Tyr	
						390					395					
AAG	GAA	CAG	CAA	AAA	CAT	GCT	CTG	AAA	ACC	CTT	GCG	ACT	CTC	TTT	TCC	1308
Lys	Glu	Gln	Gln	Lys	His	Gly	Leu	Lys	Thr	Leu	Gly	Thr	Leu	Phe	Ser	
					405					410					415	
TCA	ATG	ATG	TTC	CCA	GAT	CGA	GCT	CCT	GAT	GAC	CAA	TAT	TGA	TAT	ACA	1356
Ser	Met	Met	Phe	Pro	Asp	Arg	Ala	Pro	Asp	Asp	Gln	Tyr	Leu	Tyr	Thr	
				420					425					430		
ACA	TTT	GTT	GCG	GCT	AGC	CAC	AAT	AGA	GAT	CTT	GCT	GGA	GCT	CCA	ACG	1404
Thr	Phe	Val	Gly	Gly	Ser	His	Asn	Arg	Asp	Leu	Ala	Gly	Ala	Pro	Thr	
				435				440					445			
TCT	ATT	CTG	AAA	CAA	CCT	GTG	ACC	TCT	GAC	CTT	AAA	AAA	CTC	TTG	GCC	1452
Ser	Ile	Leu	Lys	Gln	Leu	Val	Thr	Ser	Asp	Leu	Lys	Lys	Leu	Leu	Gly	
				450			455					460				
GTA	GAG	GCG	CAA	CCA	ACT	TTT	GTG	AAG	CAT	GTA	TAC	TGG	GGA	AAT	GCT	1500
Val	Glu	Gly	Gln	Pro	Thr	Phe	Val	Lys	His	Val	Tyr	Trp	Gly	Asn	Ala	
				465			470				475					
TTT	CCT	TTG	TAT	GCC	CAT	GAT	TAT	AGT	TCT	GTA	TTG	GAA	GCT	ATA	GAA	1548
Phe	Pro	Leu	Tyr	Gly	His	Asp	Tyr	Ser	Ser	Val	Leu	Glu	Ala	Ile	Glu	

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480          495          510          525
AAG ATG GAG AAA AAC CTT GCA GGG TTC TTC TAC GCA GGA AAT AAC AAG      1596
Lys Met Glu Lys Asn Leu Pro Gly Phe Phe Tyr Ala Gly Asn Ser Lys
          300          305          310

GAT GGG CTT GCT GTT GGA AAT GTT ATA GCT TCA GGA ACC AAG GCT GCT      1644
Asp Gly Leu Ala Val Gly Ser Val Ile Ala Ser Gly Ser Lys Ala Ala
          315          320          325

GAC CTT GCA ATC TCA TAT CTT GAA TCT CAC ACC AAG CMT AAT AAT TCA      1692
Asp Leu Ala Ile Ser Tyr Leu Glu Ser His Thr Lys His Asn Asn Ser
          330          335          340

CAT TGAAGTGC TGAAGTATCC TCTAGCAATT GTGCAAAAT TCTCCCAATT      1745
His
          345

CATGTACAGT AGAAGCGAT GCGTTGCAAT TTCGAACAT CTCACCTCT TCAAGATTA      1801
ACCGTTGCTT GAGATCCAC GAGAAAGTA GTCACTATG TACTGAGAA ATGAGATTA      1849
AAACTATTA TGGGCGCGA AATGTTCTT TTTGTTTTC TCACAGTGG CTTAGCACC      1923
TTGATGTTG AAATACATT AAATTGTTG AATTGTTGA GACACATCC GTGACGTGA      1983
ATATTGCTT ATTGTGATT TACAGTACT GTTGCCAGA TTATGCTTA CCGTTTAAA      2043
AAAAAAAAA AAAAAA      2061
  
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(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 544 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

```

Met Leu Ala Leu Thr Ala Ser Ala Ser Ser Ala Ser Ser His Pro Tyr
  1              5              10              15

Arg His Ala Ser Ala His Thr Arg Arg Pro Arg Leu Arg Ala Val Leu
          20          25          30

Ala Met Ala Gly Ser Asp Asp Pro Arg Ala Ala Pro Ala Arg Ser Val
          35          40          45

Ala Val Val Gly Ala Gly Val Ser Gly Leu Ala Ala Ala Tyr Arg Leu
          50          55          60

Arg Glu Ser Gly Val Asn Val Thr Val Phe Glu Ala Ala Asp Arg Ala
          65          70          75          80

Gly Gly Lys Ile Arg Thr Asn Ser Glu Gly Gly Phe Val Trp Asp Glu
          85          90          95

Gly Ala Asn Thr Met Thr Glu Gly Glu Trp Glu Ala Ser Arg Leu Ile
          100          105          110
  
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Asp Asp Leu Gly Leu Gln Asp Lys Gln Gln Tyr Pro Asn Ser Gln His
 115 120 125
 Lys Arg Tyr Ile Val Lys Asp Gly Ala Pro Ala Leu Ile Pro Ser Asp
 130 135 140
 Pro Ile Ser Leu Met Lys Ser Ser Val Leu Ser Thr Lys Ser Lys Ile
 145 150 155 160
 Ala Leu Phe Phe Glu Pro Phe Leu Tyr Lys Lys Ala Asn Thr Arg Asn
 165 170 175
 Ser Gly Lys Val Ser Glu Gln His Leu Ser Glu Ser Val Gly Ser Phe
 180 185 190
 Cys Glu Arg His Phe Gly Arg Gln Val Val Asp Tyr Phe Val Asp Pro
 195 200 205
 Phe Val Ala Gly Thr Ser Ala Gly Asp Pro Glu Ser Leu Ser Ile Arg
 210 215 220
 His Ala Phe Pro Ala Leu Trp Asn Leu Glu Arg Lys Tyr Gly Ser Val
 225 230 235 240
 Ile Val Gly Ala Ile Leu Ser Lys Leu Ala Ala Lys Gly Asp Pro Val
 245 250 255
 Lys Thr Arg His Asp Ser Ser Gly Lys Arg Arg Asn Arg Arg Val Ser
 260 265 270
 Phe Ser Phe His Gly Gly Met Gln Ser Leu Ile Asn Ala Leu His Asn
 275 280 285
 Glu Val Gly Asp Asp Asn Val Lys Leu Gly Thr Glu Val Leu Ser Leu
 290 295 300
 Ala Cys Thr Phe Asp Gly Val Pro Ala Leu Gly Arg Trp Ser Ile Ser
 305 310 315 320
 Val Asp Ser Lys Asp Ser Gly Asp Lys Asp Leu Ala Ser Asn Gln Thr
 325 330 335
 Phe Asp Ala Val Ile Met Thr Ala Pro Leu Ser Asn Val Arg Arg Met
 340 345 350
 Lys Phe Thr Lys Gly Gly Ala Pro Val Val Leu Asp Phe Leu Pro Lys
 355 360 365
 Met Asp Tyr Leu Pro Leu Ser Leu Met Val Thr Ala Phe Lys Lys Asp
 370 375 380
 Asp Val Lys Lys Pro Leu Glu Gly Phe Gly Val Leu Ile Pro Tyr Lys
 385 390 395 400
 Glu Gln Gln Lys His Gly Leu Lys Thr Leu Gly Thr Leu Phe Ser Ser
 405 410 415
 Met Met Phe Pro Asp Arg Ala Pro Asp Asp Gln Tyr Leu Tyr Thr Thr
 420 425 430
 Phe Val Gly Gly Ser His Asn Arg Asp Leu Ala Gly Ala Pro Thr Ser
 435 440 445

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11  Leu Lys Gln Leu Val Thr Ser Asp Leu Lys Lys Leu Leu Gly Val
    450                      455                      460
Glu Gly Gln Pro Thr Phe Val Lys His Val Tyr Trp Gly Asn Ala Phe
465                      470                      475                      480
Pro Leu Tyr Gly His Asp Tyr Ser Ser Val Leu Glu Ala Ile Glu Lys
    485                      490                      495
Met Glu Lys Asn Leu Pro Gly Phe Phe Tyr Ala Gly Asn Ser Lys Asp
    500                      505                      510
Gly Leu Ala Val Gly Ser Val Ile Ala Ser Gly Ser Lys Ala Ala Asp
    515                      520                      525
Leu Ala Ile Ser Tyr Leu Glu Ser His Thr Lys His Asn Asn Ser His
    530                      535                      540

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(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1811 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 3..1589
- (D) OTHER INFORMATION: /product= "wheat protein-1 cDNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

```

GC  ACA  ACA  ATG  GGC  ACC  GGC  ACC  GTC  GCG  GCG  GCG  TCG  CCG  CTC  GGC      47
Ala  Thr  Met  Ala  Thr  Ala  Thr  Val  Ala  Ala  Ala  Ser  Pro  Leu  Arg
   1                      5                      10                      15

GCG  AGC  GTC  ACC  GCG  CCC  CCA  CAC  GCG  GTC  CCG  CCG  CGT  TCG  GCT  ACC      95
Gly  Arg  Val  Thr  Gly  Arg  Pro  His  Arg  Val  Arg  Pro  Arg  Cys  Ala  Thr
                20                      25                      30

GCG  AGC  AGC  GCG  ACC  GAG  ACT  CCG  GCG  GCG  CCC  GGC  GTC  CCG  CTG  TCC      143
Ala  Ser  Ser  Ala  Thr  Glu  Thr  Pro  Ala  Ala  Pro  Gly  Val  Arg  Leu  Ser
                35                      40                      45

GCG  GAA  TGC  GTC  ATT  GTC  GGC  GCC  GGC  ATC  AGC  GGC  CTC  TGC  ACC  GCG      191
Ala  Glu  Cys  Val  Ile  Val  Gly  Ala  Gly  Ile  Ser  Gly  Leu  Cys  Thr  Ala
                50                      55                      60

CAG  GCG  CTG  GCC  ACC  CGA  TAC  GGC  GTC  AGC  GAC  CTG  CTC  GTC  ACC  GAG      239
Gln  Ala  Leu  Ala  Thr  Arg  Tyr  Gly  Val  Ser  Asp  Leu  Leu  Val  Thr  Glu
                65                      70                      75

GCC  CCG  GAC  CCG  CCG  GGC  GGC  AAC  ATC  ACC  ACC  GTC  GAG  CGT  CCC  GAC      287

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Ala	Arg	Asp	Arg	Pro	Gly	Gly	Asn	Il	Thr	Thr	Val	Glu	Arg	Pro	Asp	
80					85				90						95	
GAG	GAG	TAC	CTG	TGG	GAG	GAG	GGA	CCC	AAC	ACC	TTC	CAG	CCC	TCC	GAC	335
Glu	Gly	Tyr	Leu	Trp	Glu	Glu	Gly	Pro	Asn	Ser	Phe	Gln	Pro	Ser	Asp	
			100					105						110		
CCG	CTC	CTC	ACC	ATG	GCC	GTG	GAC	AAC	GCG	CTC	AAG	GAT	GAC	TTC	GTG	383
Pro	Val	Leu	Thr	Met	Ala	Val	Asp	Ser	Gly	Leu	Lys	Asp	Asp	Leu	Val	
			115				120					125				
TTC	GCG	GAC	CCC	AAC	GCG	CCC	GCG	TTC	GTG	CTG	TGG	GAG	GCG	AAG	CTG	431
Phe	Gly	Asp	Pro	Asn	Ala	Pro	Arg	Phe	Val	Leu	Trp	Glu	Gly	Lys	Leu	
		130					135					140				
AGG	CCG	GTG	CCG	TGG	AAG	CCA	GCG	GAC	CTG	GCT	TTC	TTC	AAC	CTC	ATG	479
Arg	Pro	Val	Pro	Ser	Lys	Pro	Gly	Asp	Leu	Pro	Phe	Phe	Ser	Leu	Met	
		145				150					155					
AGT	ATC	CCT	GCG	AAG	CTC	AGG	GCC	GCG	CTT	GCG	GCG	CTC	GCG	ATT	CAC	527
Ser	Ile	Pro	Gly	Lys	Leu	Arg	Ala	Gly	Leu	Gly	Ala	Leu	Gly	Ile	Arg	
160					165				170					175		
CCA	CCT	CCT	CCA	GCG	CAC	GAG	GAG	TGG	GTG	GAG	GAG	TTT	GTG	CCG	CCG	575
Pro	Pro	Pro	Pro	Gly	Arg	Glu	Glu	Ser	Val	Glu	Glu	Phe	Val	Arg	Arg	
				180				185						190		
AAC	CTC	GCT	GCC	GAG	GTG	TTT	GAG	GCG	CTC	ATC	GAG	CCT	TTC	TGC	TCA	623
Asn	Leu	Gly	Ala	Glu	Val	Phe	Glu	Arg	Leu	Ile	Glu	Pro	Phe	Cys	Ser	
			195				200					205				
GGT	GTA	TAT	GCT	GCT	GAT	CCT	TGG	AAG	CTT	AGT	ATG	AAG	GCT	GCA	TTT	671
Gly	Val		Ala	Gly	Asp	Pro	Ser	Lys	Leu	Ser	Met	Lys	Ala	Ala	Phe	
		210				215						220				
GCG	AAG	GTG	TGG	AGG	TTG	GAG	GAG	ATT	GGA	GCT	AGT	ATT	ATT	GCT	GGA	719
Gly	Lys	Val	Trp	Arg	Leu	Glu	Glu	Ile	Gly	Gly	Ser	Ile	Ile	Gly	Gly	
		225				230					235					
ACC	ATC	AAG	GCG	ATT	CAG	GAT	AAA	GCG	AAG	AAC	GCT	AAA	CCG	CCA	AGG	767
Thr	Ile	Lys	Ala	Ile	Gln	Asp	Lys	Gly	Lys	Asn	Pro	Lys	Pro	Pro	Arg	
240					245					250					255	
GAT	CCC	GGA	CTT	CCG	GCA	CCA	AAG	GGA	CAG	ACG	GTG	GCA	TCT	TTC	AGG	815
Asp	Pro	Arg	Leu	Pro	Ala	Pro	Lys	Gly	Gln	Thr	Val	Ala	Ser	Phe	Arg	
				260					265					270		
AAG	GCT	CTA	GCC	ATG	CTC	CCG	AAT	GCC	ATC	GCA	TCT	AGG	CTG	GCT	AGT	863
Lys	Gly	Leu	Ala	Met	Leu	Pro	Asn	Ala	Ile	Ala	Ser	Arg	Leu	Gly	Ser	
			275				280						285			
AAA	GTC	AAG	CTG	TCA	TGG	AAG	CTT	ACG	AAC	ATT	ACA	AAG	GCG	GAC	AAC	911
Lys	Val	Lys	Leu	Ser	Trp	Lys	Leu	Thr	Ser	Ile	Thr	Lys	Ala	Asp	Asn	
		290					295					300				
CAA	GGA	TAT	GTA	TTA	GCT	TAT	GAA	ACA	CCA	GAA	GGA	CTT	GTT	TCA	GTG	959
Gln	Gly	Tyr	Val	Leu	Gly	Tyr	Glu	Thr	Pro	Glu	Gly	Leu	Val	Ser	Val	
		305				310					315					
CAG	GCT	AAA	AGT	GTT	ATC	ATG	ACC	ATC	CCG	TCA	TAT	GTT	GCT	AGT	GAT	1007
Gln	Ala	Lys	Ser	Val	Ile	Met	Thr	Ile	Pro	Ser	Tyr	Val	Ala	Ser	Asp	
320					325					330					335	

ATC TTG CCG CCA CTT TCA ATT GAT GCA GCA GAT GCA CTC TCA AAA TTC	1099
Ile Leu Arg Pro Leu Ser Ile Asp Ala Ala Asp Ala Leu Ser Lys Phe	340 345 350
TAT TAT CCG CCA GTT GCT GCT GTA ACT GTT TCA TAT CCA AAA GAA GCT	1103
Tyr Tyr Pro Pro Val Ala Ala Val Thr Val Ser Tyr Pro Lys Glu Ala	355 360 365
ATT AGA AAA GAA TGC TTA ATT GAT GGG GAG CTC CAG GGT TTC GCG CAG	1151
Ile Arg Lys Glu Cys Leu Ile Asp Gly Glu Leu Gln Gly Phe Gly Glu	370 375 380
TTG CAT CCA COT AGC CAA GGA GTC GAG ACT TTA GGG ACA ATA TAT AGC	1199
Leu His Pro Arg Ser Gln Gly Val Glu Thr Leu Gly Thr Ile Tyr Ser	385 390 395
TCT TCT CTC TTT COT AAT COT GCT GCT GCT GCA AGA GTG TTA CTT CTG	1247
Ser Ser Leu Phe Pro Asn Arg Ala Pro Ala Gly Arg Val Leu Leu Leu	400 405 410 415
AAC TAT ATC GGG GGT TCT ACA AAT ACA GGG ATC CTC TCC AAG ACT GAG	1295
Asn Tyr Ile Gly Ser Thr Asn Thr Gly Ile Val Ser Lys Thr Glu	420 425 430
AGT GAC TTA GTA GGA GCC GTT GAC COT GAC CTC AGA AAA ATG TTG ATA	1343
Ser Asp Leu Val Gly Ala Val Asp Arg Asp Leu Arg Lys Met Leu Ile	435 440 445
AAC CTT AGA GCA GCA GAC CTT TTA GCA TTA GGG GTT CCA GTG TGG CCA	1391
Asn Pro Arg Ala Ala Asp Pro Leu Ala Leu Gly Val Arg Val Trp Pro	450 455 460
CAA GCA ATA CCA CAG TTT TTG ATT GGG CAC CTT GAT CCG CTT GCT GCT	1439
Gln Ala Ile Pro Gln Phe Leu Ile Gly His Leu Asp Arg Leu Ala Ala	465 470 475
GCA AAA TCT GCA CTC GGC CAA GGC GGC TAC GAC GGG TTG TTC CTA GGA	1487
Ala Lys Ser Ala Leu Gly Gln Gly Gly Tyr Asp Gly Leu Phe Leu Gly	480 485 490 495
GGA AAC TAC GTC GCA GGA GTT GCC TTG GGC GGA TGC ATC CAG GGT GCG	1535
Gly Asn Tyr Val Ala Gly Val Ala Leu Gly Arg Cys Ile Glu Gly Ala	500 505 510
TAC GAG AGT GCC TCA CAA GTA TCT GAC TTC TTG ACC AAG TAT GCG TAC	1583
Tyr Glu Ser Ala Ser Gln Val Ser Asp Phe Leu Thr Lys Tyr Ala Tyr	515 520 525
AAG TGA TGAAGTAGT GCATCTCTTC ATTTCTTTTC ATATACGAGG TGAGGCTAGG	1639
Lys	
ATCGTAAAA CATCATGAGA TTCTGTAGTG TTCTTTAAT TGAAAAACA AATTTRAGTG	1699
ATGCATATG TGCTCTTTTC TGTAGTTGGA GCATGTACAT CGGTATGGGA TAAAGTAGAA	1759
TAGCTATTC TGCAAAAGCA GTGATTTTTT TTGAAAAAA AAAAAAAA AA	1811

(2) INFORMATION FOR SEQ ID NO:10:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 528 amino acids

43

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

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Ala Thr Met Ala Thr Ala Thr Val Ala Ala Ser Pro Leu Arg Gly
 1           5           10           15
Arg Val Thr Gly Arg Pro His Arg Val Arg Pro Arg Cys Ala Thr Ala
 20           25           30
Ser Ser Ala Thr Glu Thr Pro Ala Ala Pro Gly Val Arg Leu Ser Ala
 35           40           45
Glu Cys Val Ile Val Gly Ala Gly Ile Ser Gly Leu Cys Thr Ala Gln
 50           55           60
Ala Leu Ala Thr Arg Tyr Gly Val Ser Asp Leu Leu Val Thr Glu Ala
 65           70           75           80
Arg Asp Arg Pro Gly Gly Asn Ile Thr Thr Val Glu Arg Pro Asp Glu
 85           90           95
Gly Tyr Leu Trp Glu Glu Gly Pro Asn Ser Phe Gln Pro Ser Asp Pro
100           105           110
Val Leu Thr Met Ala Val Asp Ser Gly Leu Lys Asp Asp Leu Val Phe
115           120           125
Gly Asp Pro Asn Ala Pro Arg Phe Val Leu Trp Glu Gly Lys Leu Arg
130           135           140
Pro Val Pro Ser Lys Pro Gly Asp Leu Pro Phe Phe Ser Leu Met Ser
145           150           155           160
Ile Pro Gly Lys Leu Arg Ala Gly Leu Gly Ala Leu Gly Ile Arg Pro
165           170           175
Pro Pro Pro Gly Arg Glu Glu Ser Val Glu Glu Phe Val Arg Arg Asn
180           185           190
Leu Gly Ala Glu Val Phe Glu Arg Leu Ile Glu Pro Phe Cys Ser Gly
195           200           205
Val Tyr Ala Gly Asp Pro Ser Lys Leu Ser Met Lys Ala Ala Phe Gly
210           215           220
Lys Val Trp Arg Leu Glu Glu Ile Gly Gly Ser Ile Ile Gly Gly Thr
225           230           235           240
Ile Lys Ala Ile Gln Asp Lys Gly Lys Asn Pro Lys Pro Pro Arg Asp
245           250           255
Pro Arg Leu Pro Ala Pro Lys Gly Gln Thr Val Ala Ser Phe Arg Lys
260           265           270
Gly Leu Ala Met Leu Pro Asn Ala Ile Ala Ser Arg Leu Gly Ser Lys
275           280           285
Val Lys Leu Ser Trp Lys Leu Thr Ser Ile Thr Lys Ala Asp Asn Gln
290           295           300

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Gly Tyr Val Leu Gly Tyr Glu Thr Pro Glu Gly Leu Val Ser Val Gln
305 310 315 320
Al Lys Ser Val Ile Met Thr Ile Pro Ser Tyr Val Ala Ser Asp Ile
325 330 335
Leu Arg Pro Leu Ser Ile Asp Ala Ala Asp Ala Leu Ser Lys Phe Tyr
340 345 350
Tyr Pro Pro Val Ala Ala Val Thr Val Ser Tyr Pro Lys Glu Ala Ile
355 360 365
Arg Lys Glu Cys Leu Ile Asp Gly Glu Leu Gln Gly Phe Gly Gln Leu
370 375 380
His Pro Arg Ser Glu Gly Val Glu Thr Leu Gly Thr Ile Tyr Ser Ser
385 390 395 400
Ser Leu Phe Pro Asn Arg Ala Pro Ala Gly Arg Val Leu Leu Leu Asn
405 410 415
Tyr Ile Gly Gly Ser Thr Asn Thr Gly Ile Val Ser Lys Thr Glu Ser
420 425 430
Asp Leu Val Gly Ala Val Asp Arg Asp Leu Arg Lys Met Leu Ile Asn
435 440 445
Pro Arg Ala Ala Asp Pro Leu Ala Leu Gly Val Arg Val Trp Pro Gln
450 455 460
Ala Ile Pro Gln Phe Leu Ile Gly His Leu Asp Arg Leu Ala Ala Ala
465 470 475 480
Lys Ser Ala Leu Gly Gln Gly Gly Tyr Asp Gly Leu Phe Leu Gly Gly
485 490 495
Asn Tyr Val Ala Gly Val Ala Leu Gly Arg Cys Ile Glu Gly Ala Tyr
500 505 510
Glu Ser Ala Ser Gln Val Ser Asp Phe Leu Thr Lys Tyr Ala Tyr Lys
515 520 525

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1847 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(ix) FEATURES:

- (A) NAME/KEY: CDS
- (B) LOCATION: 55..1683
- (D) OTHER INFORMATION: /product= "soybean preox-1 cDNA"

(ml) SEQUENCE DESCRIPTION: SEQ ID NO:11:

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TTTAGCACA GTGTGAAGA TAACAAACGA ATAGTCCAT TACTGTAAAC AACCT ATG      57
Met
1

GTT TCC GTC TTC AAC GAG ATC CTA TTC CCG CCG AAC CAA ACC CTT CTT      105
Val Ser Val Phe Asn Glu Ile Leu Phe Pro Pro Asn Glu Thr Leu Leu
5 10 15

CCG CCG TCC CTC CAT TCC CCA ACC TCT TTC TTC ACC TCT CCG ACT CCA      153
Arg Pro Ser Leu His Ser Pro Thr Ser Phe Phe Thr Ser Pro Thr Arg
20 25 30

AAA TTC CCT CCG TCT CCG CCT AAC CCT ATT CTA CCG TCC TCC ATT CCG      201
Lys Phe Pro Arg Ser Arg Pro Asn Pro Ile Leu Arg Cys Ser Ile Ala
35 40 45

GAG GAA TCC ACC CCG TCT CCG CCC AAA ACC AGA GAC TCC CCG CCG GTG      249
Glu Glu Ser Thr Ala Ser Pro Pro Lys Thr Arg Asp Ser Ala Pro Val
50 55 60 65

GAC TCC GTC GTC GTC GCG GGA GCG GTC ACC GCG CTC TCC ATC GCG CAG      297
Asp Cys Val Val Val Gly Gly Gly Val Ser Gly Leu Cys Ile Ala Gln
70 75 80

GCC CTC GCG ACC AAA CAC GCC AAT GCC AAC GTC GTC GTC ACC GAG GCC      345
Ala Leu Ala Thr Lys His Ala Asn Ala Asn Val Val Val Thr Glu Ala
85 90 95

CGA GAC CCG GTC GCG GCG AAC ATC ACC ACC ATG GAG AGG GAC GGA TAC      393
Arg Asp Arg Val Gly Gly Asn Ile Thr Thr Met Glu Arg Asp Gly Tyr
100 105 110

CTC TCG GAA GAA GCG CCC AAC AGC TTC CAG CCT TCT GAT CCA ATG CTC      441
Leu Trp Glu Glu Gly Pro Asn Ser Phe Gln Pro Ser Asp Pro Met Leu
115 120 125

ACC ATG GTG GTG GAC AGT GGT TTA AAG GAT GAG CTT GTT TTG GCG GAT      489
Thr Met Val Val Asp Ser Gly Leu Lys Asp Glu Leu Val Leu Gly Asp
130 135 140 145

CCT GAT GCA CCT CCG TTT GTC TTG TCG AAC AGG AAG TTG AGG CCG GTG      537
Pro Asp Ala Pro Arg Phe Val Leu Trp Asn Arg Lys Leu Arg Pro Val
150 155 160

CCG CCG AAG CTC ACT GAT TTG CCT TTC TTT GAC TTG ATG AGC ATT GGT      585
Pro Gly Lys Leu Thr Asp Leu Pro Phe Phe Asp Leu Met Ser Ile Gly
165 170 175

GCG AAA ATC AGG GCT GCG TTT GGT CCG CTT GGA ATT CCG CCT CCT CCT      633
Gly Lys Ile Arg Ala Gly Phe Gly Ala Leu Gly Ile Arg Pro Pro Pro
180 185 190

CCA GGT CAT GAG GAA TCG GTT GAA GAG TTT GTT CGT CCG AAC CTT GGT      681
Pro Gly His Glu Glu Ser Val Glu Glu Phe Val Arg Arg Asn Leu Gly
195 200 205

GAT GAG GTT TTT GAA CCG TTG ATA GAG CCT TTT TGT TCA GCG GTC TAT      729
Asp Glu Val Phe Glu Arg Leu Ile Glu Pro Phe Cys Ser Gly Val Tyr
210 215 220 225

GCA GCG GAT CCT TCA AAA TTA AGT ATG AAA GCA GCA TTC GCG AAA GTT      777

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Ala Gly Asp Pro Ser Lys Leu Ser Met Lys Ala Ala Phe Gly Lys Val	
230 239 240	
TGG AAG CTG GAA AAA AAT GGT GGT AGC ATT ATT GGT GGA ACT TTC AAA	825
Trp Lys Leu Glu Lys Asn Gly Gly Ser Ile Ile Gly Gly Thr Phe Lys	
345 256 255	
GCA ATA CAA GAG AGA AAT GGA GGT TCA AAA CCA CCT CGA GGT CCG GGT	873
Ala Ile Gln Glu Arg Asn Gly Ala Ser Lys Pro Pro Arg Asp Pro Arg	
260 265 270	
CTG CCA AAA CCA AAA GGT CAG ACT GTT GGA TCT TTC CCG AAG GGA GTT	921
Leu Pro Lys Pro Lys Gly Thr Val Gly Ser Phe Arg Lys Gly Leu	
275 280 285	
ACC ATG TTG CTT GAT GCA ATT TCT GCC AGA CTA GGC AAC AAA GTA AAG	969
Thr Met Leu Pro Asp Ala Ile Ser Ala Arg Leu Gly Asn Lys Val Lys	
290 295 300 305	
TTA TCT TGG AAG CTT TCA AGT ATT AGT AAA CTG GGT AGT GGA GAG TAC	1017
Leu Ser Trp Lys Leu Ser Ser Ile Ser Lys Leu Asp Ser Gly Glu Tyr	
310 315 320	
AGT TTG ACA TAT GAA ACA CCA GAA GGA GTG GTT TCT TTG CAG TGC AAA	1065
Ser Leu Thr Tyr Glu Thr Pro Glu Gly Val Val Ser Leu Glu Cys Lys	
325 330 335	
ACT GTT GTC CTG ACC ATT CCT TCC TAT GTT GGT AGT ACA TTG CTG GGT	1113
Thr Val Val Leu Thr Ile Pro Ser Tyr Val Ala Ser Thr Leu Leu Arg	
340 345 350	
CCT CTG TCT GCT GCT GCT GCA GAT GCA CTT TCA AAG TTT TAT TAC CCT	1161
Pro Leu Ser Ala Ala Ala Ala Asp Ala Leu Ser Phe Thr Tyr Tyr Pro	
355 360 365	
GCA GTT GGT GCA GTT TCC ATA TCC TAT CCA AAA GAA GCT ATT AGA TCA	1209
Pro Val Ala Ala Val Ser Ile Ser Tyr Pro Lys Glu Ala Ile Arg Ser	
370 375 380 385	
GAA TGC TTG ATA GAT GGT GAG TTG AAG GGG TTT GGT CAA TTG CAT CCA	1257
Glu Cys Leu Ile Asp Gly Glu Leu Lys Gly Phe Gly Gln Leu His Pro	
390 395 400	
GGT AGC CAA GGA GTG GAA ACA TTA GGA ACT ATA TAC AGC TCA TCA CTA	1305
Arg Ser Gln Gly Val Glu Thr Leu Gly Thr Ile Tyr Ser Ser Ser Leu	
405 410 415	
TTC CCG AAC GGA GCA CCA CCT GGA AAG GTT CTA CTC TTG AAT TAC ATT	1353
Phe Pro Asn Arg Ala Pro Pro Gly Arg Val Leu Leu Leu Asn Tyr Ile	
420 425 430	
GGA GGA GCA ACT AAT ACT GGA ATT TTA TCG AAG ACU GAC AGT GAA CTT	1401
Gly Gly Ala Thr Asn Thr Gly Ile Leu Ser Lys Thr Asp Ser Glu Leu	
435 440 445	
GTG GAA ACA GTT GAT CCA GAT TTG AAG AAA ATC CTT ATA AAC CCA AAT	1449
Val Glu Thr Val Asp Arg Asp Leu Arg Lys Ile Leu Ile Asn Pro Asn	
450 455 460 465	
GCC CAG GAT CCA TTT GTA GTG GGG GTG AGA CTG TCG CTT CAA CTT ATT	1497
Ala Gln Asp Pro Phe Val Val Gly Val Arg Leu Trp Pro Gln Ala Ile	
470 475 480	

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CCA CAG TTC TTA GTT GGC CAT CTT GAT CTT CTA GAT GTT GCT AAA GCT      1545
Pro Glu Phe Leu Val Gly His Leu Asp Leu Leu Asp Val Ala Lys Ala
485 490 495

TCT ATC AGA AAT ACT GGG TTT GAA GGG CTC TTC CTT GGG GGT AAT TAT      1595
Ser Ile Arg Asn Thr Gly Phe Glu Gly Leu Phe Leu Gly Gly Asn Tyr
500 505 510

GTG TCT GGT GTT GGC PRO GGA GGA TGC GTT GAG GGA GGC TAT GAG GTA      1645
Val Ser Gly Val Ala Leu Gly Arg Cys Val Glu Gly Ala Tyr Glu Val
515 520 525

CCA GCT GAA GTA AAC GAT TTT CTC ACA AAT AGA GTG TAC AAA      1695
Ala Ala Glu Val Asn Asp Phe Leu Thr Asn Arg Val Tyr Lys
530 535 540

TAGTAGCAAT TTTTGTITT GTGTGTGAAT GGTGTGTGGG ACTGTCTGTGT TCCATTGAT      1745
TATATATATG TGAAGGTTTC TCAAAATTCGT TCAATAGGTT TTTGGGGCT TCTATTCGTG      1805
ATAATGTAAA ATCTCTTGA AGTTTGAAA AAAAAAAAAA AAAA      1865

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(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 543 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID 12:

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Met Val Ser Val Phe Asn Glu Ile Leu Phe Pro Pro Asn Gln Thr Leu      1
1 5 10 15

Leu Arg Pro Ser Leu His Ser Pro Thr Ser Phe Phe Thr Ser Pro Thr      20
20 25 30

Arg Lys Phe Pro Arg Ser Arg Pro Asn Pro Ile Leu Arg Cys Ser Ile      35
35 40 45

Ala Glu Glu Ser Thr Ala Ser Pro Pro Lys Thr Arg Asp Ser Ala Pro      50
50 55 60

Val Asp Cys Val Val Val Gly Gly Gly Val Ser Gly Leu Cys Ile Ala      65
65 70 75 80

Gln Ala Leu Ala Thr Lys His Ala Asn Ala Asn Val Val Val Thr Glu      85
85 90 95

Ala Arg Asp Arg Val Gly Gly Asn Ile Thr Thr Met Glu Arg Asp Gly      100
100 105 110

Tyr Leu Trp Glu Glu Gly Pro Asn Ser Phe Gln Pro Ser Asp Pro Met      115
115 120 125

Leu Thr Met Val Val Asp Ser Gly Leu Lys Asp Glu Leu Val Leu Gly      130
130 135 140

Asp Pro Asp Ala Pro Arg Phe Val Leu Trp Asn Arg Lys Leu Arg Pro      145
145 150 155 160

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Val Pro Gly Lys Leu Thr Asp Leu Pro Phe Phe Asp Leu Met Ser Ile
165 170 175

Gly Gly Lys Ile Arg Ala Gly Phe Gly Ala Leu Gly Ile Arg Pro Pro
180 185 190

Pro Pro Gly His Glu Glu Ser Val Glu Glu Phe Val Arg Arg Asn Leu
195 200 205

Gly Asp Glu Val Phe Glu Arg Leu Ile Glu Pro Phe Cys Ser Gly Val
210 215 220

Tyr Ala Gly Asp Pro Ser Lys Leu Ser Met Lys Ala Ala Phe Gly Lys
225 230 235 240

Val Trp Lys Leu Glu Lys Asn Gly Gly Ser Ile Ile Gly Gly Thr Phe
245 250 255

Lys Ala Ile Glu Glu Arg Asn Gly Ala Ser Lys Pro Pro Arg Asp Pro
260 265 270

Arg Leu Pro Lys Pro Lys Gly Glu Thr Val Gly Ser Phe Arg Lys Gly
275 280 285

Leu Thr Met Leu Pro Asp Ala Ile Ser Ala Arg Leu Gly Asn Lys Val
290 295 300

Lys Leu Ser Trp Lys Leu Ser Ser Ile Ser Lys Leu Asp Ser Gly Glu
305 310 315 320

Tyr Ser Leu Thr Tyr Glu Thr Pro Glu Gly Val Val Ser Leu Glu Cys
325 330 335

Lys Thr Val Val Leu Thr Ile Pro Ser Tyr Val Ala Ser Thr Leu Leu
340 345 350

Arg Pro Leu Ser Ala Ala Ala Ala Asp Ala Leu Ser Lys Phe Tyr Tyr
355 360 365

Pro Pro Val Ala Ala Val Ser Ile Ser Tyr Pro Lys Glu Ala Ile Arg
370 375 380

Ser Glu Cys Leu Ile Asp Gly Glu Leu Lys Gly Phe Gly Glu Leu His
385 390 395 400

Pro Arg Ser Glu Gly Val Glu Thr Leu Gly Thr Ile Tyr Ser Ser Ser
405 410 415

Leu Phe Pro Asn Arg Ala Pro Pro Gly Arg Val Leu Leu Leu Asn Tyr
420 425 430

Ile Gly Gly Ala Thr Asn Thr Gly Ile Leu Ser Lys Thr Asp Ser Glu
435 440 445

Leu Val Glu Thr Val Asp Arg Asp Leu Arg Lys Ile Leu Ile Asn Pro
450 455 460

Asn Ala Glu Asp Pro Phe Val Val Gly Val Arg Leu Trp Pro Glu Ala
465 470 475 480

Ile Pro Glu Phe Leu Val Gly His Leu Asp Leu Leu Asp Val Ala Lys
485 490 495

Ala Ser Ile Arg Asn Thr Gly Phe Glu Gly Leu Phe Leu Gly Gly Asn
500 505 510
Tyr Val Ser Gly Val Ala Leu Gly Arg Cys Val Glu Gly Ala Tyr Glu
515 520 525
Val Ala Ala Glu Val Asn Asp Phe Leu Thr Asn Arg Val Tyr Lys
530 535 540

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 583 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

(A) NAME/KEY: promoter
(B) LOCATION: 1..583
(D) OTHER INFORMATION: /function= "arabidopsis protein-1 promoter"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

GAATTCGGAT CGAATTATAT AATTATCAT AATTGGAATA AGCATGTTTC CTTTATTAA	60
AGAGGTTTAA TAAAGTTTGC TAATAATGGA CTTTGACTTC AACTCGATT CTCATGTAA	120
TAATTAATAT TTACATCAA ATTGCTCAC TAATATACC AATTAACAT ACTAAATGT	180
TAATTCGCAA ATAAACACT AATTCGAAT AAGGCTCAT TATGATAAC ACATATTGA	240
CTTGATAAG CAAAGCAAAA ATAATGGTT TCAAGTTTTC GATTATATAT GACAAAAAA	300
AAAAAGGTT TGATTATATA TCTATTGGC CTATACCAT GTATACAAA TTGAGGCTA	360
ACTAAATTA TAAATAAAC GTAATGCTC TTTTATATT TGGTCAAC CCACCTCTA	420
ACCAACCA AGAAAAAT ATACGCTAG GTACACAGAC TATGCTGTC TGTGATTGA	480
GCTGAATAT TCTGCTGTC TTCTCTTC TTCTGAAGAA GATTACCAA TCTGAAGAA	540
ACCAAGAGC TGACAAAAT CGAATTCTC TGGATTTC ATG	583

The invention as described herein is contemplated to include the following enumerated embodiments:

1. A recombinant DNA molecule comprising a plant protoporphyrinogen oxidase (protox) promoter or a functionally equivalent derivative thereof.
2. A chimeric gene comprising a plant protox promoter operably linked to a heterologous DNA coding sequence.
3. The chimeric gene of claim 2 wherein said plant protox promoter is from a protox-1 gene.
4. The chimeric gene of claim 2 wherein said plant protox promoter is from a protox-2 gene.
5. The chimeric gene of claim 2 wherein said protox promoter is from a plant selected from the group consisting of *Arabidopsis*, soybeans, cotton, tobacco, sugar beet, oilseed rape, maize, wheat, sorghum, rye, oats, turf grass and rice.
6. The chimeric gene of claim 5 wherein said promoter is from an *Arabidopsis* plant.
7. The chimeric gene of claim 6 wherein said promoter is at least 300 nucleotides in length.
8. The chimeric gene of claim 7 wherein said promoter is at least 500 nucleotides in length.

9. The chimeric gene of claim 8 wherein said promoter has the sequence set forth in SEQ ID No. 13.

10. The chimeric gene of claim 2 wherein said heterologous coding sequence encodes a modified, herbicide-resistant form of a plant enzyme.

11. The chimeric gene of claim 10 wherein said plant enzyme is selected from the group consisting of isochlorogenic acid phosphatase (ICPD), EPSP synthase, glutamine synthetase (GS), acetyl coenzyme A carboxylase, nucleotidyl transferase, and glutathione transferase (GST).

12. The chimeric gene of claim 11 wherein said plant enzyme is GST.

13. A recombinant DNA vector comprising the recombinant DNA molecule of claim 1.

14. Plant tissue comprising the chimeric gene of claim 2.

15. A plant comprising the chimeric gene of claim 2.

16. The plant of claim 15 wherein said plant is selected from the group consisting of Arabidopsis, soybean, cotton, tobacco, sugar beet, oilseed rape, maize, wheat, sorghum, rye, oats, turf grass and rice.

ABSTRACT OF DISCLOSURE

5 Promoters naturally associated with plant protoporphyrinogen oxidase (protoporphyrinogen oxidase) coding sequences, and derivatives thereof, are provided. These promoters can be used to control the expression of an operably linked heterologous coding sequence in a plant cell. These promoters are particularly useful for expressing modified forms of herbicide target enzymes, particularly modified forms of protoporphyrinogen oxidase, to achieve tolerance to herbicides which inhibit the corresponding
10 unmodified enzymes. Recombinant DNA molecules and chimeric genes comprising these promoters are provided, as well as plant tissue and plants containing such chimeric genes.